

Green Tea Extract Induces Apoptosis in the AGS Gastric Carcinoma Cell Line

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Abstract

Green tea has long been thought to possess anticancer properties. Additionally, the polyphenol components of green tea have demonstrated ability to inhibit 26S proteasome function. As there have been reports describing varied responses of gastric carcinoma cells to green tea treatment, the role of green tea treatment on proteasome function in the gastric carcinoma cell line AGS was investigated. Presented here are findings demonstrating that green tea extract is capable of inhibiting proteasome function of AGS cells. Furthermore, treatment of AGS cells with the green tea polyphenol (-)-Epigallocatechin-3-gallate resulted in a similar level of proteasome inhibition. It was also discovered in this study that proteasome inhibition in AGS cells resulted in a buildup of Kip1/p27 and $\text{I}\kappa\text{B}\alpha$, proteins that are involved in the progression through the G1/S checkpoint during cell division. Proteasome inhibition by (-)-Epigallocatechin-3-gallate led to induction of apoptosis of AGS cells. Our results described here strongly suggest that green tea consumption is capable of inducing programmed cell death in gastric carcinoma cells through inhibition of proteasome activity. It should be noted, however, that consumption of green tea during anti-cancer protocols has been reported to reduce the effectiveness of a specific subset of chemotherapeutic compounds. In summary, consumption of green tea in gastric carcinoma patients may be effective in targeting cancer cells and slowing the progression of gastric cancers. Future studies of these natural products may provide some structural information which will allow for the development of the next generation of anti-cancer chemotherapeutics.

Keywords: Green tea; Proteasome; Gastric carcinoma; Apoptosis

Introduction

Green tea has long been consumed worldwide, and numerous reports have provided evidence of the beneficial nutritional aspects of green tea [1]. More specifically, epidemiological studies have provided correlative evidence that consumption of green tea reduces cancer development [2]. Additionally, green tea extracts have been found to possess anti-tumor activity in animal models and cultured cells [3,4]. Effects on tumors and cancer cells have been attributed to the polyphenol compounds contained within green tea extracts [5-7]. The catechin (-)-Epigallocatechin-3-gallate (EGCG) is thought to be the most potent of the polyphenol components, and has been shown to mediate an induction of an apoptotic cascade in certain tumor cells [8-10]. Varying degrees of tumor cell inhibition have been reported, however, depending both on cell lines studied [11] and also which catechins were used as inhibitors [12].

Nam, et al. [13] demonstrated that the polyphenols were capable of inhibiting the activity of the 26S proteasome complex. The 26S proteasome is a large multi-protein complex that regulates many cellular metabolic processes through protein degradation of ubiquitinated target proteins [14]. Processes ranging from protein sorting [15] to peptide presentation in immune system functions [16] are regulated by proteasome function. Recent reports have also demonstrated that the proteasome is a dynamic complex that influences processes such as activated transcription [17-20] and DNA repair [21-23] solely through the action of a non-proteolytic proteasome sub-complex known as the 19S regulatory complex. Interestingly, among other proteasome-regulated cellular processes are apoptosis and progression through the cell division cycle [24,25]. Cyclin Dependent Kinases (CDK) inhibitor proteins such as Kip1/p27 must be degraded by the proteasome in order for the cell to proceed through the G1/S checkpoint [26]. Failure to degrade these proteins in a temporal fashion has been shown to trigger apoptosis in a variety of cell types [26]. Progression from mitosis also relies on the temporal degradation of M-phase promoting cyclins [24], thus demonstrating the importance for proper proteasome function in "normal" cellular function.

Inhibition of proteasome activity has long been thought to be an attractive method by which to target cancer cells. The compound bortezomib (Velcade, Millennium Pharmaceuticals) was the first of such chemical proteasome inhibitors to be approved as an anti-cancer treatment [27]. Bortezomib therapy demonstrated effectiveness against multiple myeloma, but there are recent reports of association of bortezomib treatment and toxicity, as well as tumor cell types exhibiting resistance to the inhibitor [27]. The success of bortezomib has sparked an interest in the study of proteasome inhibitors as anti-cancer chemotherapeutics.

In this work, the effects of green tea extract and the polyphenol EGCG were studied in their ability to affect the growth of the AGS gastric adenocarcinoma cell line. It was found that incubation of AGS cells with either green tea extract or pure EGCG inhibited proteasome function, and led to the buildup of ubiquitinated proteins to a similar extent as treatment of AGS cells with the proteasome inhibitor MG-132. Use of purified human proteasome complexes further confirmed the inhibitory properties of EGCG. To evaluate cellular effects of EGCG-mediated proteasome inhibition, induction of apoptosis was studied. EGCG was found to induce apoptosis, and also to lead to a buildup of CDK inhibitor proteins involved in the progression from G1 to S phase. Taken together these results suggest that EGCG, through its proteasome inhibiting properties, is capable of inducing apoptosis of AGS cells via cell cycle blockage at the G1/S border.

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Materials and Methods

Cell Line and Culture Conditions

The human gastric carcinoma cell line AGS, culture medium, and supplements were obtained from American Type Culture Collection (ATCC, Manassas, VA) unless otherwise noted. Cells were cultured in Ham's F12K medium supplemented with 2 mM final concentration L-glutamine and 10% Fetal Bovine Serum (Atlanta Biologicals, Flowery Branch, GA). Cultures were maintained at 37°C and 5% CO₂ during all experiments.

Treatment of Gastric Carcinoma Cells with Green Tea Extract, EGCG, or the Proteasome Inhibitor MG-132

For experiments that incorporated incubation of cells with chemicals, 1×10^6 cells were plated in a total of 3 ml culture medium in 6 well culture plates. Each condition was plated in triplicate. Green tea extract (Pharmanex, Provo, UT) and epigallocatechin gallate (EGCG; Sigma-Aldrich, St. Louis, MO) were resuspended in sterile water, and the 26S proteasome inhibitor Z-Leu-Leu-Leu-CHO (MG-132, Enzo Life Sciences, Farmingdale, NY) was resuspended in dimethyl sulfoxide (DMSO; Sigma-Aldrich, St. Louis, MO). Inhibitors were incubated at the indicated concentrations for 1 hour prior to cell harvesting. Following the 1 hour time period, cells were washed twice in 1X Dulbecco's PBS (Life Technologies, Carlsbad, CA) and pellets were stored at -80°C. Whole cell extracts (WCE) were generated from cell pellets using the M-PER protein extraction reagent (Pierce Biotechnology, Rockford, IL) and protein concentrations were determined by bicinchoninic acid (BCA) protein quantitation (Pierce Biotechnology, Rockford, IL) on a spectrophotometer at 562 nm.

26S Proteasome Assays

Activity of the 26S proteasome was assayed by measuring the chymotryptic activity of the complex. Briefly, 50 µg WCE was pre-warmed to 37°C along with 25 µl assay buffer (50mM Tris-HCl, pH 7.5). Sterile water was used to bring the final reaction volume to 100 µl. The reaction was initiated upon the addition of 5 µl of 0.5 mM Succinyl-Leucine-Leucine-Valine-Tyrosine-7-Amido-4-methylcoumarin (Succ-LLVY-AMC; Bachem, Torrance, CA). Reactions were performed in triplicate and allowed to progress at 37°C for 2 hours. Fluorescence of liberated AMC corresponded to proteasomal peptidase activity and was measured in a BioTek Synergy2 microplate reader (BioTek US, Winooski, VT), using excitation wavelength of 340 nm and emission wavelength of 465 nm.

For assays that tested *in vitro* inhibition of purified human 26S proteasome, the proteasome complex was obtained from Boston Biochem (Cambridge, MA). Purified proteasome was added to a final concentration of 10 nM to the assay components as described above. Green tea extract, EGCG, or MG-132 were added to intact proteasome complexes, and allowed to incubate on ice for 15 minutes prior to adding the fluorogenic substrate Succ-LLVY-AMC. Enzyme activity was measured as described above, and was reported as Relative Fluorescence Units (RFU).

Western Blot Analysis

Whole cell extracts generated from mock or inhibitor treated cell populations were used in western blot analyses. Fifty micrograms of total protein for each condition was loaded in SDS-PAGE gels and transferred to nitrocellulose membranes (BIORAD, Hercules, CA). Accumulation of ubiquitinated proteins as a result of proteasome inhibition was observed using a primary antibody raised against

ubiquitin (Boston Biochem, Cambridge, MA). Antisera raised against poly ADP ribose polymerase-1 (PARP-1), IκBα, Kip1/p27 and β-actin were obtained from EMD Millipore (Billerica, MA). Secondary antibodies conjugated to horseradish peroxidase were obtained from Pierce (Rockford, IL). Bands corresponding to the proteins of interest were visualized on autoradiography film (Phenix Research Products, Candler, NC) using the Supersignal Pico Western Blot Detection kit (Pierce Biotechnology, Rockford, IL).

Caspase 3 Assay

Induction of apoptosis was addressed by measuring the activation of caspase 3 in whole cell extracts either mock treated or treated with EGCG at various concentrations. Caspase 3 activity corresponded to the cleavage of the caspase 3 specific substrate Ac-Aspartic acid-Glutamic acid-Valine-Aspartic acid-para-nitro aniline (Ac-DEVD-pNA; EMD Millipore, Billerica, MA). Briefly, 200 µM Ac-DEVD-pNA was incubated with 25 µg total protein in 100 µl buffer consisting of 50 mM HEPES, pH 7.4, 100 mM NaCl, 0.1% CHAPS, 10 mM DTT, 0.1 mM EDTA and 10% glycerol. Reactions were allowed to proceed for 2 hours at 37°C, and 96 well plates were read at 405 nm on a BioTek Synergy2 microplate reader (BioTek US, Winooski, VT). Reactions were each performed in triplicate, and the data reported represents the mean of measurements.

Data Analysis

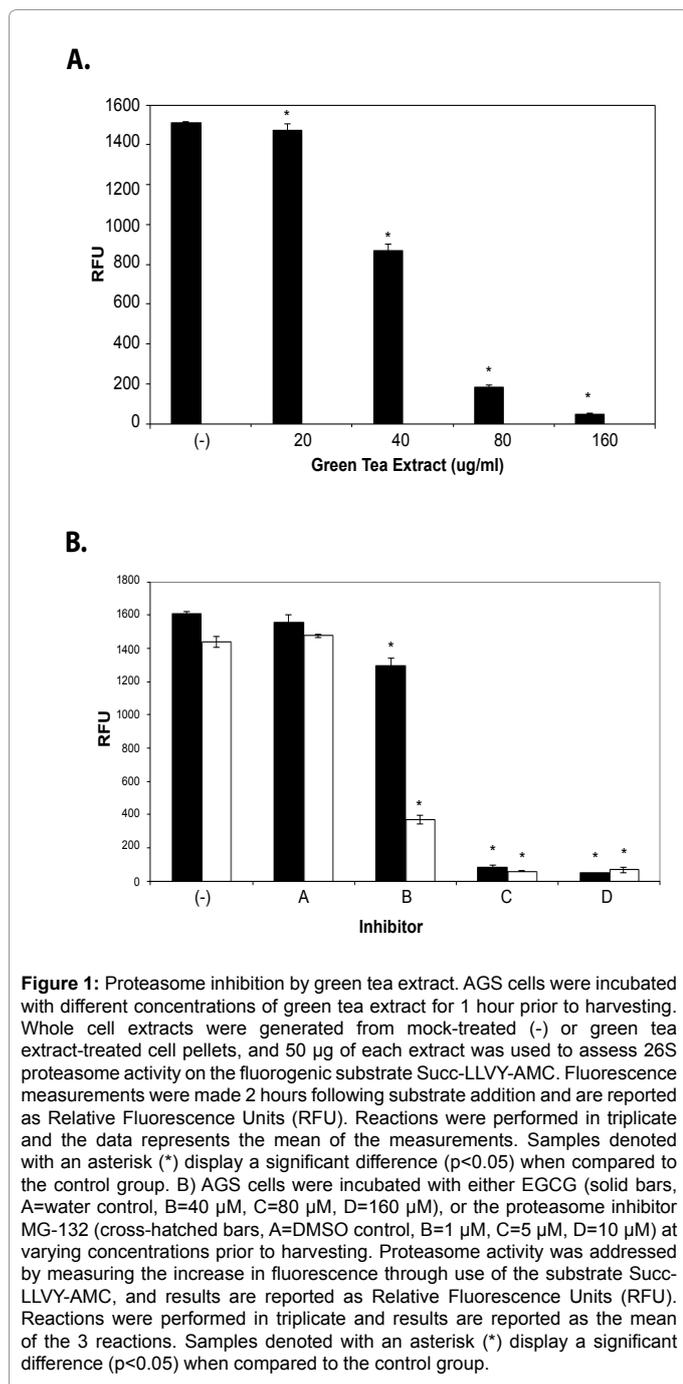
Data was analyzed for significance using a one-way ANOVA and Dunnett's (2-sided) test in IBM SPSS. A p-value of less than 0.05 was considered to be significant.

Results

Green tea extract inhibits 26S activity in AGS cells

Green and black tea extracts are known to possess polyphenol compounds that resemble traditional proteasome inhibitors such as β-lactone [13]. Using Jurkat T cells, Nam, et al [13] demonstrated that polyphenol components of green tea extracts possess significant proteasome inhibitory properties. Horie, et al [28] however presented data that demonstrated a gradient of effects by green tea catechins on different types of cultured cells. AGS cells were incubated with increasing concentrations of green tea extract in order to assess cellular responses to green tea extracts. Whole cell extracts from mock or green tea treated cells were subsequently generated and used in an assay of 26S chymotrypsin-like proteasome activity. As shown in Figure 1A, incubation of AGS cells with crude green tea extract resulted in a concentration-dependent inhibition of the chymotrypsin-like activity of the proteasome. Interestingly, although there was no significant proteasome inhibition in cells treated with 20 µg/ml green tea extract, exposure of AGS cells to 80 µg/ml green tea extract resulted in approximately a 90% inhibition of 26S proteasome activity.

The major anti-cancer compounds found in green tea extracts include the polyphenol catechin compounds, such as (-) epigallocatechin-3-gallate (EGCG; [29]). To address the ability of EGCG to specifically inhibit proteasome activity, AGS cells were incubated with different concentrations of pure EGCG (Figure 1B, black bars). As seen with cells exposed to green tea extract, cells treated with 80-160 µM EGCG exhibited significant levels of proteasome inhibition. Cells mock treated with water did not noticeably differ in the extent of AGS cell proteasome inhibition. As a source of comparison, AGS cells treated with the proteasome specific inhibitor MG-132 (Figure 1B, cross-hatched bars) demonstrated a similar pattern of inhibition of proteasomal chymotrypsin-like activity.



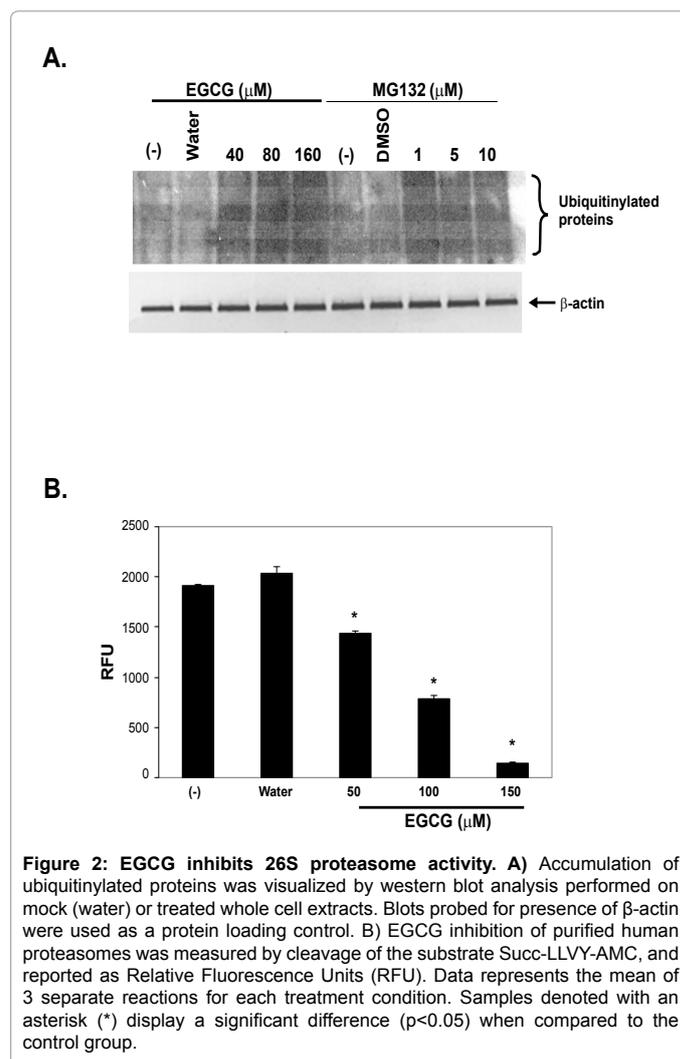
As the assay described above measures peptidase activity against a synthetic substrate, it was imperative to understand the influence of EGCG on proteasome function in AGS cells. Inhibition of proteasome function is known to result in the buildup of ubiquitinated proteins in a whole cell extract [30]. Cells treated with either EGCG or MG-132 was assessed for the relative quantity of ubiquitinated proteins found in response to chemical treatment. As shown in Figure 2A, western blot analysis using an anti-ubiquitin primary antibody demonstrated a characteristic “smear” in untreated (-) or mock-treated (A) cell extracts. Incubation of AGS cells with increasing concentrations of EGCG demonstrated an increasing intensity of signal of ubiquitinated proteins. Of interest was the finding that

effects of EGCG treatment of AGS cells mimicked the cells treated with a specific chemical proteasome inhibitor (MG-132). Taken together, the results presented in Figures 1B and 2A demonstrate that EGCG inhibits gastric carcinoma cell proteasome function to an extent similar to the inhibition patterns of MG-132.

In order to specifically address the proteasome inhibitory properties of EGCG, a purified 26S proteasome complex was used in an assay as described above. Data shown in Figure 2B demonstrates that highly purified human 26S proteasomes are also readily inhibited by EGCG. This data further suggests that the effects of green tea catechins function to target the proteasome complex.

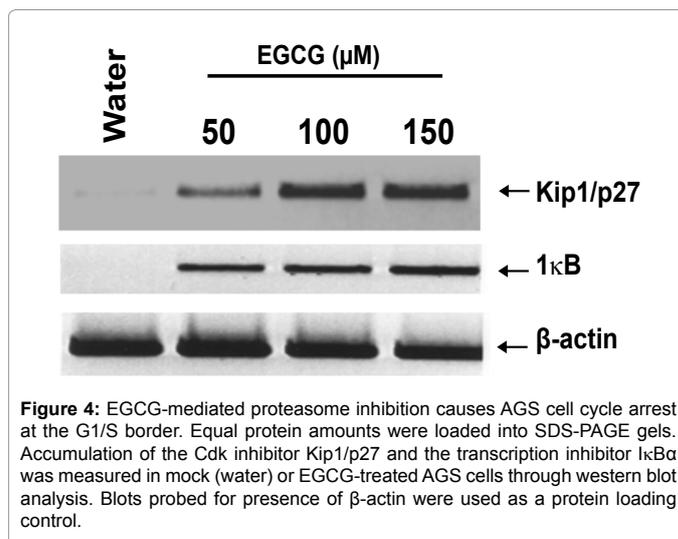
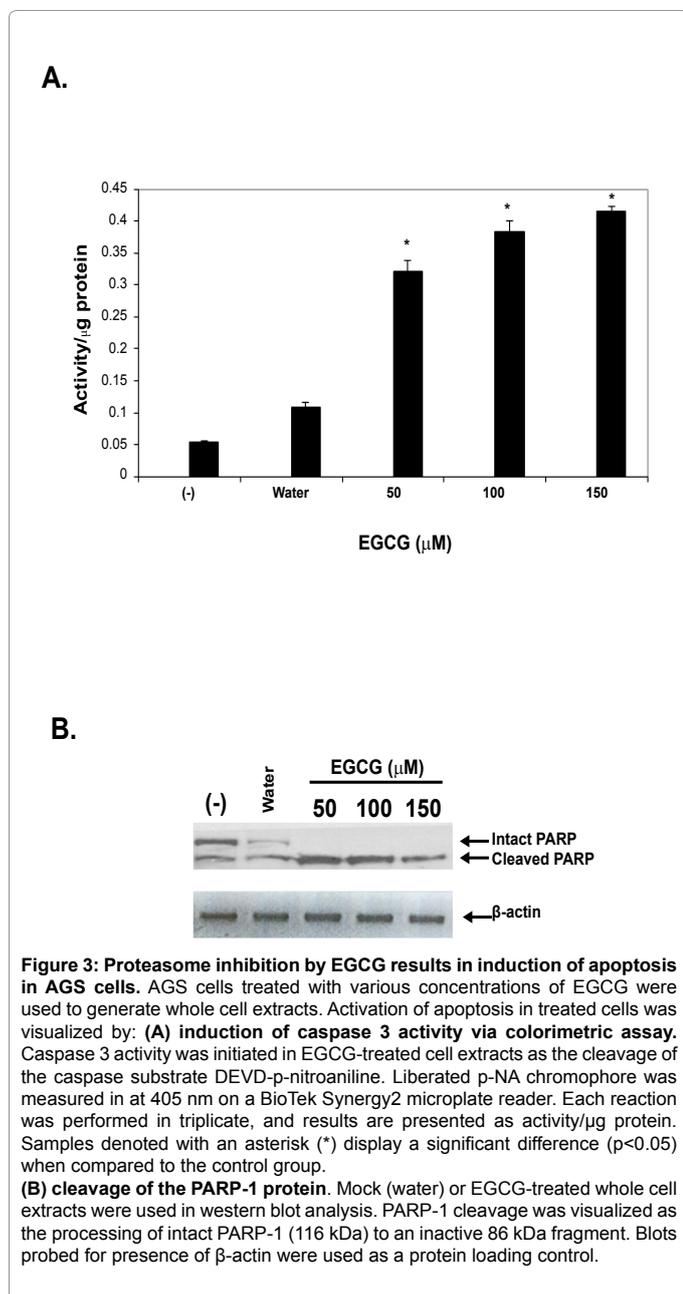
Proteasome inhibition in AGS cells results in induction of apoptosis

Proteasome inhibition is known to efficiently induce apoptosis in cultured cells [30-32]. Additionally, anti-cancer chemotherapeutics such as bortezomib function to slow cancer cell growth through inhibition of proteasome activity. Based on the fact that both green tea extract and EGCG were capable of marked proteasome inhibition in cultured gastric carcinoma cells (AGS), it was hypothesized that proteasome inhibition would result in the induction of apoptosis in AGS cells. Whole cell extracts from AGS cells treated with EGCG concentrations known to inhibit proteasome activity were used in



an assay for enzymatic activity of caspase 3, a protein involved in the apoptotic pathway [33]. Data shown in Figure 3A demonstrates that incubation of AGS cells with increasing concentrations of EGCG resulted in the induction of caspase 3 activity.

To further address induction apoptosis by EGCG in AGS cells, whole cell extracts described above were used as the input proteins in western blot analyses. Blots were assayed for the presence of cleaved poly ADP ribose polymerase-1 (PARP-1), an indicator of irreversible, late stage apoptotic events [34,35]. As shown in Figure 3B, treatment of AGS cells with EGCG resulted in the cleavage of intact PARP-1 protein, as evidenced by the lack of detectable intact PARP-1 protein. Taken together, the results strongly suggest that proteasome inhibition in AGS cells by EGCG is capable of inducing programmed cell death of these cells.



EGCG treated AGS cells are blocked at the G1/S border of the cell cycle

Traditional chemical inhibition of proteasomes results in the arrest of cells at the G1/S checkpoint of the cell cycle. To further investigate the mechanistic role of EGCG-mediated proteasome inhibition in AGS cells, western blot analysis was performed in which 2 proteins involved in progression of cells through the G1/S border were investigated. Accumulation of the cyclin dependent kinase inhibitor Kip1/p27 and transcription inhibitor protein $\text{I}\kappa\text{B}\alpha$ are hallmarks of cell cycle arrest at the G1/S checkpoint [36-38]. As evidenced in Figure 4, relative protein amounts of Kip1/p27 and $\text{I}\kappa\text{B}\alpha$ were markedly increased in response to EGCG treatment, even at the lowest concentration of EGCG used experimentally (50 μM). Little to no detectable Kip1/p27 or $\text{I}\kappa\text{B}\alpha$ was observed in the mock treated samples (water). Additionally, antibodies raised against β -actin were used as a protein loading control. Results presented in Figure 4 strongly suggest that AGS cells exposed to EGCG are arrested at the G1/S border as a result of proteasome inhibition.

Discussion

Presented here is experimental evidence to support the long-standing idea that the chemical components of green tea possess anti-cancer cell properties. Using the gastric carcinoma cell line AGS, it was found that green tea extract was capable of inhibiting enzymatic activity of the 26S proteasome. Coincidentally, a similar extent of proteasome inhibition was observed following treatment of AGS cells (and purified proteasomes) with the green tea polyphenol component (-)-Epigallocatechin-3-gallate (EGCG). Caspase activity assays provided evidence of apoptotic induction following exposure of AGS cells to EGCG. Furthermore, it was observed that EGCG mediated proteasome inhibition resulted in the accumulation of cell division cycle proteins involved in the release of cells from the G1/S border.

Epidemiological studies have provided correlative evidence to describe the association between green tea consumption and growth inhibition of tumors [1-4]. Subsequent work has attributed the majority of the anti-cancer effects of green tea to the polyphenol catechins found in green tea [39-41]. Furthermore, Nam, et al. [13] determined that concentrations of green tea catechins responsible for these effects lie close to, or at physiological levels found in green tea extracts. There are many proposed cellular effects following exposure to EGCG, including acting as an anti-oxidant in the scavenging of free oxygen radicals as

well as acting to inhibit angiogenesis [42]. Interestingly, the proposed effect of polyphenols on regulation of the cell cycle machinery [24,25] may play the biggest role in inhibition of cancer progression.

Dysregulation of the machinery involved in cell division is known to induce the formation of cancer cells [32], and temporal degradation of proteins is necessary for proper progression through the cell division cycle [32]. Regulation of the cell division cycle proteins is largely a function of the 26S proteasome, a multi subunit complex responsible for the majority of the non-lysosomal protein degradation in eukaryotes [14]. Coincidentally, EGCG of green tea was found to potently inhibit the chymotrypsin-like enzymatic activity of the proteasome complex [13]. Consistent with this idea are the results found in this study. Both green tea extract and EGCG were found to significantly inhibit the proteasomal chymotrypsin-like activity in the AGS gastric carcinoma cell line. This inhibition reached levels similar to the well-characterized proteasome inhibitor MG-132 (Figure 1B). Of note in the work presented here is that a general accumulation of ubiquitinated proteins was also found in AGS cells exposed to EGCG (Figure 2A). When taken together, our results suggest that EGCG treatment of AGS cells greatly reduces the overall proteasome function.

Green tea and EGCG induce apoptotic pathways in a variety of cell lines originally isolated from gastric tumors [28,42]. Horie et al [28] however presented evidence to demonstrate that different gastric carcinoma cell lines exhibited varied susceptibility to EGCG. Synthetic EGCG analog compounds have also demonstrated varied proteasome-inhibiting ability [12,43]. Results described in the present study demonstrate that inhibition of the proteasome complex induces cell cycle arrest at the G1/S border (Figure 4) and programmed cell death of AGS cells (Figure 3), suggesting that EGCG treatment of AGS cells mimics the global effects of proteasome inhibition by traditional inhibitor chemicals [30].

Proteasome inhibition has become an exciting target for the development of novel anti-cancer chemotherapeutics. The proteasome inhibitor bortezomib (Velcade, Millennium Pharmaceuticals) was approved in 2003 as a treatment option for multiple myeloma, and greatly increased the interest and research in the field of proteasome inhibition. Currently, bortezomib is being investigated for its ability to treat other types of cancers [44]. With respect to the findings reported here that EGCG results in apoptosis of gastric carcinoma cells, one might assume that a combination of bortezomib and EGCG (or green tea) would work synergistically to inhibit cancer cell growth. Recently however, Golden et al. [44] demonstrated that consumption of green tea unexpectedly reduces the therapeutic activity of bortezomib and other boronic acid-based inhibitors (MG-262 and PS-IX). Results described here and elsewhere underscore the need for a more thorough understanding of not only the interactions of potential inhibitor compounds with each other, but also the knowledge regarding the detailed responses of various cell types to proteasome inhibition.

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