

Integrating Next Generation Sequencing, Bioinformatics and Cytogenomics in the Study of Brazilian Mammals

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Abstract

Since the discovery of the DNA double helix, understanding the complexity and diversity of genomes has been a major focus of genetics, including medical and evolutionary research. Sequencing methods have been developed since the 1970's, starting with the first-generation or Sanger sequencing.

Keywords: Next generation sequencing; Bioinformatics; Cytogenomics; Sanger sequencing; DNA-DNA hybridizations

Mini Review

A second-generation or Next-Generation Sequencing (NGS) was developed in 2005 and consisted of a fast speed and high cost-benefit sequencing, when compared to traditional methodologies [1-3]. NGS symbolized a new revolution in genomics, as it has the potential to tackle biological problems related to the genome, transcriptome and epigenome of any species. Albeit the many advantages of this technology, it requires sufficient knowledge of bioinformatics to analyze the data. Moreover, it produces short reads which makes it difficult to assemble whole genomes [4]. Although a third-generation of sequencing that aims at overcoming the limitation of NGS has been evolving since 2008 [5], the second-generation is still largely employed.

Among the diversity of algorithms and bioinformatics tools for the analyses of NGS data, our group has been successfully using RepeatExplorer [6,7], implemented on the galaxy platform, to identify and characterize repetitive DNAs in species with low coverage genome sequencing (Figure 1a).

Once the sequenced data are available, RepeatExplorer can be used to perform all-to-all comparison analyses of sequence reads by similarities and to build clusters of overlapping reads that represent repetitive DNA families (Figure 1b).

This is an easy, fast and more accurate methodology to identify even low abundant repetitive families, sometimes undetected in experimental studies. In the Repeat Explorer output, we look for clusters composed of tandemly repeated satellite DNAs (satDNAs), or transposable elements, complementing the analyses with other softwares such as Tandem Repeats Finder [8] and Dotlet [9], and the databases for repetitive DNAs called Repbase [10]. The bioinformatic analyses are followed by molecular biology experiments, including localization of the repetitive DNAs on chromosome preparations by fluorescent in situ hybridization (FISH; Figure 1b).

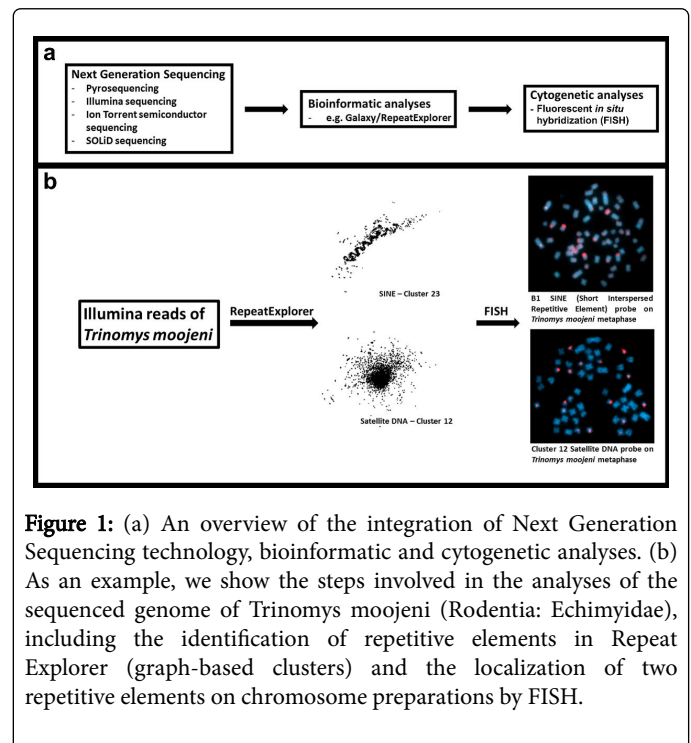


Figure 1: (a) An overview of the integration of Next Generation Sequencing technology, bioinformatic and cytogenetic analyses. (b) As an example, we show the steps involved in the analyses of the sequenced genome of *Trinomys moojeni* (Rodentia: Echimyidae), including the identification of repetitive elements in Repeat Explorer (graph-based clusters) and the localization of two repetitive elements on chromosome preparations by FISH.

We have been using this approach to address several aspects of repetitive DNAs, such as origin, structure, organization, variability, chromosome distribution, and the role of these elements in chromosome/genome evolution and their possible use as taxonomic markers. At the chromosomal level, the distribution of repetitive DNAs may be evolutionarily important, since they have been related to genome and karyotype remodeling in several eukaryotes [11].

The integration of the methodologies described above may be applied to the study of any eukaryote group, and we will use as examples the two mammalian groups in which we have concentrated in the last few years: Brazilian rodents and monkeys. Although both of these groups have been intensively studied, they still present many taxonomic problems, basic cytogenetic information is missing for many taxa, and little is known about the mechanisms related to their

genome evolution. We believe that both taxa are especially interesting for genome research, as many of their genera present highly variable karyotypes, including B-chromosomes, multiple sex chromosome systems, and several rearrangements, such as inversions, translocations, fusions/fissions, constitutive heterochromatin variation and centromere repositioning, differentiating even closely related taxa.

For example, we studied Brazilian rodent species of the genus *Trinomys* (family Echimyidae), typically difficult to identify taxonomically by their morphology and sometimes even by their karyotypes. We investigated the satDNAs of three specimens (*Trinomys moojeni*, *T. setosus*, and *T. sp.*) after sequencing their genome by NGS. A genomic library was prepared using the Nextera Kit, according to the manufacturer's instructions (Illumina Inc., San Diego, CA), and paired-end sequenced in a single flowcell using a MiSeq instrument with the MiSeq Reagent Kit V3 (600 cycles).

Sequencing achieved coverages between 9.1% and 4.5% (*T. moojeni*), 14.1% and 7% (*T. setosus*), and 16.7% and 8.3% (*T. sp.*), assuming 3 pg and 6 pg genome sizes, respectively. Although the sequence data generated had low coverage, it was enough to study the repetitive sequences of these species. Thus, after clustering Illumina reads by similarity, using RepeatExplorer, we identified a satDNA family with 350 bp motifs that showed species-specific features after sequence comparisons and phylogenetic analyses. These species have very similar karyotypes, so we concluded that, although the divergence time among them was not enough to produce karyotypic changes, it was sufficient to allow accumulate some sequence differentiation in this satDNA, which may therefore be used as a species-specific taxonomic marker (Araújo et al. in preparation).

In another work, we recently identified the CarB satDNA in the sequenced genome of the New World monkey *Callithrix jacchus* [12]. Previous studies based on genomic DNA digestions and DNA-DNA hybridizations led Alves to suggest that this satDNA was specific for *Mico* species. In fact, we found that CarB represents only 0.1% of the *C. jacchus* genome and such small amount combined to its sequence divergence might have hindered its detection in the previous experimental studies [13].

Conclusion

In summary, new technologies, like NGS, combined with bioinformatic analyses, have been fostering the identification of a large

set of repetitive DNAs that can now be used, for example, in cytogenetic and taxonomic studies. Moreover, the integration of sequenced data with physical mapping on chromosomes of highly diverse groups, like rodents and new world monkeys, promises to greatly contribute to a better understanding of several issues related to chromosome and genome structure and evolution.

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