

Neuroprotective Effects of *Aframomum melegueta* Extract after Experimental Traumatic Brain Injury

Aswathi Kumar, Deborah Kennedy-Boone, Harris A Weisz, Bridget A Capra, Tatsuo Uchida, Kristofer Jennings, Maria-Adelaide Micci, Margaret Parsley, Douglas S DeWitt, Donald S. Prough and Helen L Hellmich*

The University of Texas Medical Branch, 301 Harborside Drive, Galveston TX 77555-0830, USA

Abstract

Aframomum melegueta is an herb in the ginger family that has been shown to have anti-inflammatory, anti-oxidative, anti-diabetic and antimicrobial properties. We investigated the possibility that the seeds of this herb, which are consumed by gorillas and used as a spice in West and North African cuisine, could have neuroprotective effects in a rat model of traumatic brain injury (TBI). Using Fluoro-Jade, an anionic fluorescent stain that is a well-established marker of degenerating neurons, we found that an extract of *Aframomum*, PMI-006, significantly reduced numbers of dying, Fluoro-Jade-positive neurons in the rat hippocampus 24 hr after TBI. We used an antibody to CD11b (Ox42), a microglial marker, to show that PMI almost completely reduced microglial activation—a hallmark of injury-induced inflammation—in the rat hippocampus and cortex. To elucidate the molecular mechanisms underlying the neuroprotective effects of PMI-006, we used *RT² Profiler* pathway-focused PCR arrays representing oxidative stress, cytokine & chemokines and NFκB cell signaling pathways to interrogate PMI-induced changes in hippocampal gene expression after TBI. We found that PMI treatment ameliorated the effects of brain injury and, in several cases, restored injury-induced gene expression changes to sham control levels. PMI treatment did not significantly alter functional outcome in the Morris Water Maze, a neurobehavioral test of hippocampal-dependent spatial memory. However, because of its safety profile and because it mitigates the effects of TBI on stress and inflammatory signaling pathways that are associated with TBI pathology, PMI could be potentially beneficial in reducing neurodegeneration in TBI survivors.

Keywords: Traumatic brain injury; *Aframomum melegueta*; Hippocampus; Inflammatory genes; Oxidative stress genes; NFκB signaling genes; Fluoro-Jade

Introduction

Natural products and their derivatives were the basis for early medicines and continue to provide a rich source of drugs today, with up to 60% of approved new chemical entities (NCE) originating from natural sources [1–5]. A major reason for this is thought to be that the small molecules produced by diverse organisms have evolved to interact with biological targets which are broadly conserved across the animal and plant kingdoms, and that these compounds are generated and maintained in diverse lineages of living organisms [4].

Aframomum melegueta is a species in the ginger family, Zingiberaceae, members of which are known to possess strong anti-inflammatory and/or antibacterial properties [6,7]. In West African folk medicine, *Aframomum melegueta* seeds, known as Grains of Paradise, are valued for their warming and digestive properties as well as numerous medicinal effects. Researchers in a biotechnology company, Phytomedics, found that a derivative of this plant, PMI-006, has powerful anti-inflammatory effects, comparable to the well-known anti-inflammatory drugs Vioxx, Celebrex and Bextra but without their adverse side effects. Thus, it has been suggested that *Aframomum* might successfully be used to treat diseases with inflammation as their hallmarks, such as cardiovascular conditions, arthritis, osteoporosis and Alzheimer's disease. Anecdotal evidence also suggests that for centuries, native African healers have used *Aframomum* to treat infections of all kinds. Recent studies have provided experimental support for these antimicrobial effects [8]. Because of these medicinal properties, we reasoned that this compound may prove neuroprotective in our rat model of fluid percussion traumatic brain injury (TBI), which is a clinically relevant model for human brain injury [9–11].

TBI is a leading cause of death and lifelong disability, but there are no approved drugs for millions of TBI survivors due to the failure of all pharmacotherapeutic treatments in clinical trials [12–14]. Thus, it is imperative that we continue to explore novel sources of drug candidates for TBI. In millions of patients surviving civilian and

military trauma, the costs of TBI include disruption of daily functions, irreparable cognitive impairment, inability to return to work and overall decreased quality of life [15]. Thornhill et al. [16] evaluated 459 survivors of mild, moderate and severe TBI at one year after injury and reported that even the mildly injured patients had a 43% incidence of cognitive impairment, much of which is associated with injury-induced neurodegeneration in the hippocampus, a region in the medial temporal lobe that is critical to learning, memory and executive function [17,18]. The critical role of the hippocampus in brain function is evident in neurological disorders that are associated with cognitive dysfunction. Memory loss and dementia in Alzheimer's patients are closely correlated with loss of hippocampal neurons, and TBI patients commonly experience memory and learning deficits that are linked to hippocampal damage [19–21]. Functional neuroimaging studies have shown that the hippocampus is actively engaged during navigational tasks in humans and that hippocampal damage directly influences its interactions with other brain regions during memory retrieval [22,23].

The purpose of this study was to determine if an extract derived from *Aframomum melegueta* (PMI-006) could improve functional outcome after experimental TBI and if so, to identify underlying mechanisms of neuroprotection. Using an established fluorescent marker of degenerating neurons, Fluoro-Jade [24], immunohistochemical analysis of activated microglia, gene expression analysis using pathway-focused PCR arrays and a test of hippocampal-dependent cognitive

*Corresponding author: Helen L Hellmich, University of Texas Medical Branch, 301 Harborside Drive, Galveston TX 77555-0830, USA, Tel: 409-772-4216; Fax: 409-772-1224; E-mail: hhellmic@utmb.edu

Received November 24, 2014; Accepted December 22, 2014; Published February 01, 2015

Citation: Kumar A, Kennedy-Boone D, Weisz HA, Capra BA, Uchida T, et al. (2015) Neuroprotective Effects of *Aframomum Melegueta* Extract after Experimental Traumatic Brain Injury. Nat Prod Chem Res 3: 167. doi:10.4172/2329-6836.1000167

Copyright: © 2015 Kumar A, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

function, the Morris Water Maze [25], we characterized the neuro protective effects and therapeutic potential of this natural product-derived compound.

Materials and Methods

Preparation of *Aframomum melegueta*

PMI-006 extract was obtained from Phytomedics, Inc (Jamesburg, NJ). Preparation of the extract has been described by Ilic et al. [26]. Phytomedics scientists have used PMI 006 at doses 250-1000 mg/kg and saw physiological effects during the next 24 hours. From toxicology tests, they determined that NOAEL (no-observed-adverse-effect-level) is 1500 mg/kg. Experimental results generated at Phytomedics showed that it takes 3 hours of pretreatment for PMI-006 to be effective. All of PMI botanical drugs are administered orally. Per Phytomedics suggestions, we solubilized PMI-006 in 100% ethanol and prepared a 10% ethanol solution brought to volume with corn oil and administered it to our experimental rats via oral gavage 1 hour after injury.

Fluid percussion injury

The Institutional Animal Care and Use Committee of The University of Texas Medical Branch approved all experimental protocols. Adult, male, Charles River Sprague-Dawley rats (300-400g) were anesthetized with 4% isoflurane, intubated, then mechanically ventilated and prepared for fluid percussion TBI. A craniotomy was performed laterally to the sagittal suture, midway between the lambda and bregma structures. The fluid percussion device was then attached, and the animal was subjected to severe lateral fluid percussion traumatic brain injury (TBI) as previously described [10]. The rats were sacrificed 24hrs post-injury for neuronal counting of degenerating neurons [27] and PCR array analysis. For immunohistochemistry, rat brains were dissected out eleven days after injury following the final day of Morris Water Maze testing, immediately frozen on dry ice, and stored at -80°C.

Neuronal counting

Rats were randomly assigned to receive TBI plus 10mg, 100mg, 250mg, 500mg or 1000mg of PMI-006 (mg/kg body weight) or TBI plus vehicle alone (10% ethanol in corn oil) via oral gavage. Animals were survived for 24 Hours post injury, sacrificed, their brains removed, sectioned on a cryostat and 10 µm frozen coronal brain sections were stained with Fluoro-Jade [27] and counter stained with a nissl stain, 1% cresyl violet. A blinded investigator then counted Fluoro-Jade positive neurons in the CA1/2 and CA3 regions on the ipsilateral (injured) side of the rat hippocampus using an Olympus BX51 Fluorescent Microscope. The numbers of FJ-positive neurons were quantified for each of the treatment groups and reported as mean +/- SEM and analyzed using an analysis of variance (ANOVA) followed by the Bonferroni-Dunn test with $\alpha=0.05$. Statistical computations were carried out using PROC GLM in SAS®, Release 9.1 [28].

Neurobehavioral assessment using Morris Water Maze (MWM)

MWM procedures assessing working memory are described in detail by Hamm et al. [25]. Tank parameters for the MWM were as follows: a black tank (180 cm diameter, 28 cm depth) was filled with ambient temperature water. When filled, the tank contained a clear plastic platform hidden beneath the surface of the water. Acquisition blocks consisted of two daily trials over five consecutive days (7-11 days after injury). At the onset of each acquisition trial, rats were placed by hand in the pool facing the tank wall. There were four zones and four

starting areas for the rats; each rat was given two trials at each starting point. In this version of the Morris water maze, which assessed working memory, the location zone of the hidden platform was randomized between trials on the same day. Animals (10 sham, 8 TBI, and 12 TBI + PMI-006) were allowed to swim a maximum of 2 min to find the hidden platform and the latency was recorded for each trial. If the rat failed to find the platform after 2 min, it was placed on the platform by the experimenter. All rats were allowed to remain on the platform for 15 sec before being returned to a heated holding box for a 4 min inter trial interval. The SMART computer program (SMART program, San Diego Instruments, Inc., San Diego, CA) was used to collect, store and analyze the behavioral data. Statistical analyses were performed using PROC MIXED in SAS® (version 9.4) with no adjustment for covariance necessary and a Tukey adjustment for multiple comparisons.

Immunohistochemistry for assessment of microglial activation

To assess the effects of PMI-006 on TBI-induced inflammation, we performed immunohistochemical analysis of TBI, PMI-006 treated and sham control rat brains using an antibody to CD11b (OX-42), a marker of microglial activation. Eleven days after injury, rats were sacrificed (n=3/group), perfused with 4% paraformaldehyde, brains collected and 10 µm frozen sections were cut on a cryostat. Sections were then incubated overnight with a 1° antibody (mouse anti-CD11b; 1:2000, BD Biosciences, San Jose, CA). The following morning, sections were incubated with a 2° antibody (Alexa 594 goat anti-mouse; 1:400, Life Technologies, Grand Island, NY) at ambient temperature, and then mounted with DAPI (stains nuclei) for imaging. An Olympus BX51 Fluorescent Microscope was used to visualize the hippocampal formation and surrounding cortical regions.

RNA isolation

Total RNA was isolated from dissected ipsilateral (injured side) hippocampal tissue samples using the Ultraspec RNA isolation System (Biotecx Laboratories, Inc. Houston, TX) following the manufacturer's protocol for RNA isolation from whole tissue. Genomic DNA contamination was removed by with DNase treatment (Ambion, Austin TX) and then RNA was ethanol precipitated and brought up in nuclease-free water. RNA from TBI, TBI + PMI-006 treated, and sham injured animals was assessed for quality and quantity on an Agilent Bioanalyzer (Agilent Technologies, Santa Clara CA) with RIN values consistently averaging 7.0 to 8.0 or higher. Approximately 1 µg of each RNA sample was reverse transcribed using the RT² First Strand Kit (SA Biosciences) in preparation for use in PCR arrays.

RTProfiler PCR arrays

To profile the expression of genes related to oxidative stress, cytokines and chemokines, and NF-κB signaling pathways after TBI, TBI+PMI-006 and sham injury, quantitative real-time PCR was performed using the RT² Sybr-green Profiler PCR Arrays (SA Biosciences, Valencia, CA) following manufacturer's protocols. The expression of genes involved in the oxidative stress, cytokine & chemokine, and NF-κB signaling pathways (n=3, 4, 3/per group, respectively) in the TBI alone and TBI + PMI-006 treated rat brains were compared to the baseline levels of the same genes in the sham injured rat brains. Data analysis was based on the delta-delta CT method with normalization of the raw data to 5 housekeeping genes and calculations of the fold changes were done using the Analysis Web portal program provided by SA Biosciences.

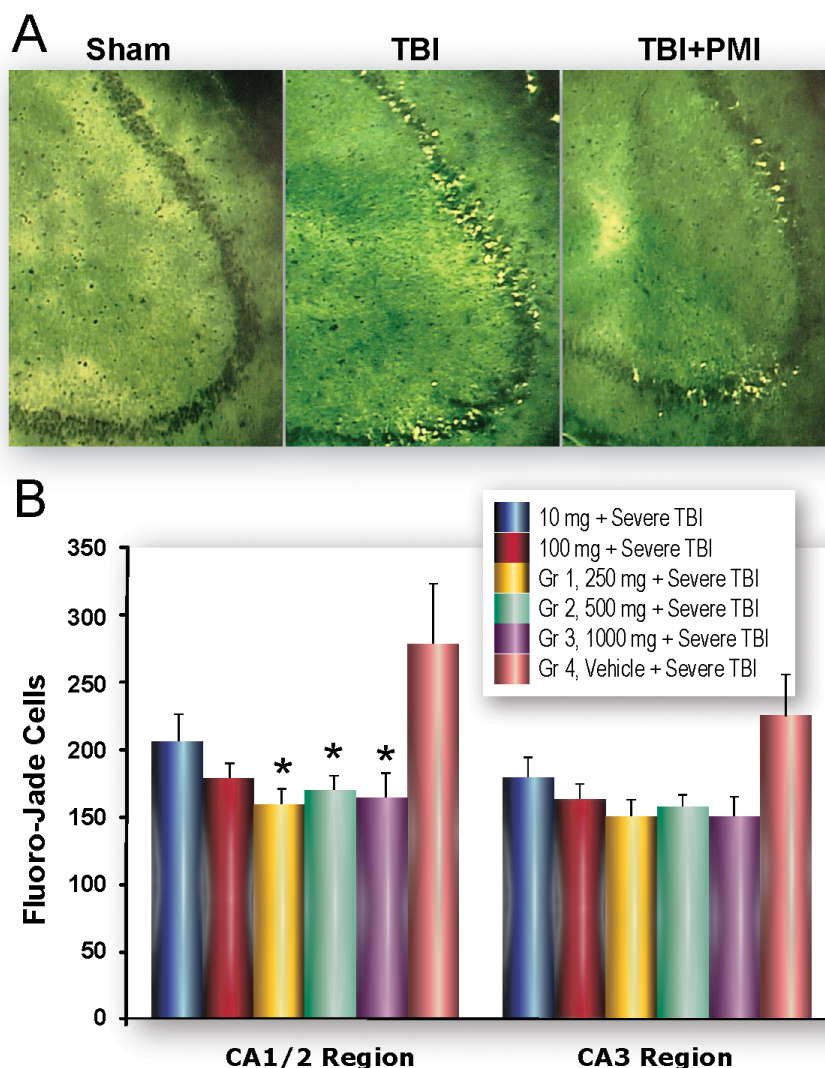


Figure 1: Effects of PMI-006, an extract of *Aframomum melegueta*, on hippocampal cell death after traumatic brain injury. (A) 10 μ m sections were taken from TBI, TBI + PMI-006, and sham injured animals, stained with Fluoro-Jade a marker for neuronal injury and then counter stained with 1% Cresyl Violet. (B) Dose response for neuroprotective effects of PMI-006 in the hippocampal subfields. Asterisks denote significant differences between numbers of Fluoro-Jade-positive neurons in severe TBI plus vehicle vs TBI + PMI treated animals; *, p value ≤ 0.05

Results

PMI-006 reduces neuronal injury in rat hippocampus

To determine the lowest dose of PMI-006 that provides maximal protective effects, we performed a dose response. In animals treated with PMI-006, there was a significant decrease in the number of Fluoro-Jade positive neurons in groups (250, 500, 1000 mg/kg) compared with vehicle treatment alone in the CA1/2 region suggesting that PMI 006 reduces neuronal injury in the hippocampus (Figure 1A, B). For Figure 1A and all subsequent experiments, we used a dose of 250mg/kg, the lowest dose which was associated with the greatest reduction in neuronal injury.

PMI-006 reduces microglial activation

Because microglial activation is a hallmark of TBI [29], we performed immunohistochemical analysis of injured rat brain sections using an antibody to CD11b, a marker of activated microglia. PMI-006

had a remarkably ameliorative effect on microglial activation after TBI (Figure 2), almost completely abolishing injury-induced inflammation in the hippocampus and cortex.

Pathway-focused PCR array analysis

The PCR super arrays (Oxidative stress, Cytokine and Chemokines, and NF κ B pathways) were chosen because of the reported anti-inflammatory and anti-oxidative properties of PMI-006. In all three PCR array data sets, we found that, as expected, severe TBI alone induced multiple proinflammatory and oxidative stress-inducing genes (complete dataset of fold changes are shown in Tables S1, S2 and S3). In each pathway-focused array, we compared significant changes induced by PMI treatment to significant changes induced by TBI alone (Figure 3). Most of these significantly affected genes are associated with injury-associated pathways that are implicated in the pathogenesis of TBI (Supplemental References). In most of these cases, we observed a consistent and reproducible trend: PMI-006 treatment reversed or normalized the effects of TBI in the direction of sham control levels. Interestingly, these

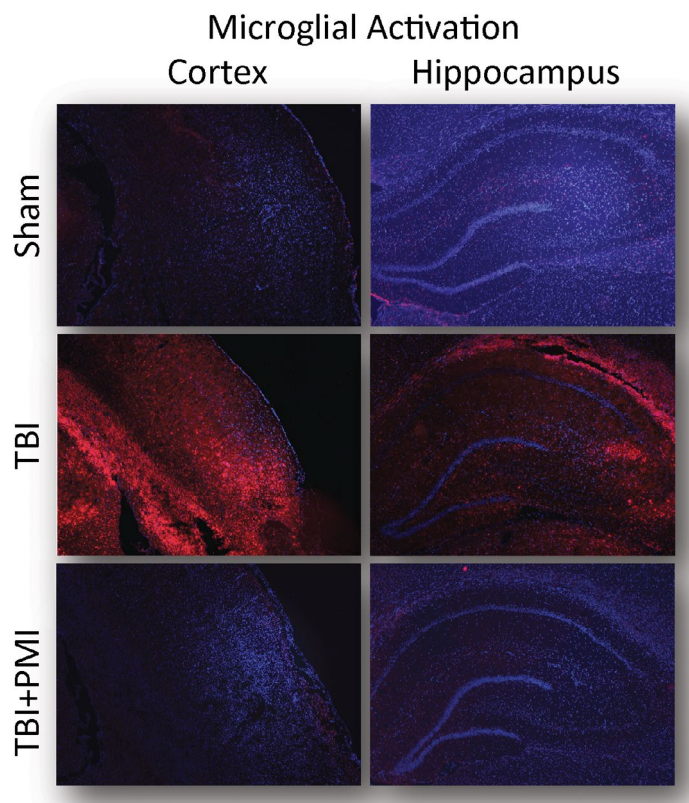


Figure 2: Effects of PMI-006 on post-injury induced microglial activation. Immunohistochemistry for OX-42 (mouse anti-CD11b; 1:2000) in the rat hippocampus and cortex shows decreased microglial activation 11 days after treatment with PMI-006. Nuclei (in blue) are stained with DAPI.

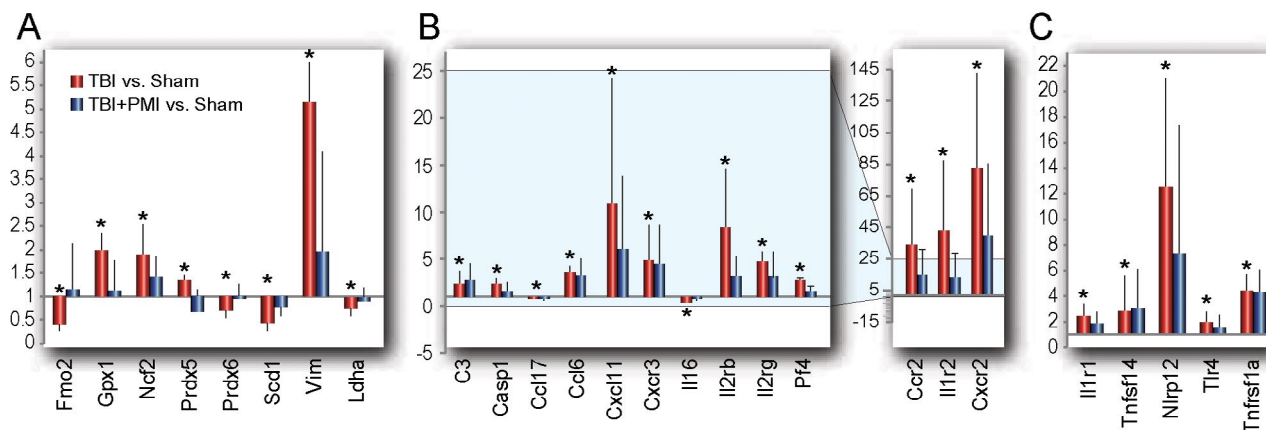


Figure 3: Effects of PMI-006 on oxidative stress, cytokine & chemokine, and NF-κB signaling PCR array genes. TBI significantly altered genes (in red vs. sham injury; *: denotes p-value ≤ 0.05) that are implicated in oxidative stress (A), cytokine & chemokine (B), and NF-κB signaling (C) pathways. Drug treatment with PMI after traumatic brain injury attenuated the TBI-induced up and down regulation of genes (in blue vs. sham injury). P-values are based on a Student's *t*-test of the replicate $2^{-(\Delta\Delta Ct)}$ values of each gene displayed with 95% confidence intervals.

gene expression results suggested that PMI-006 mitigated the effects of TBI on both deleterious genes and some protective genes, such as GPX-1, that are involved in the brain's endogenous protective responses to injury [30].

Morris water maze

We examined the effect of PMI-006 on neurobehavioral outcome after TBI using the Morris Water Maze, an Established test of

hippocampal-dependent working memory (Figure 4). There were no apparent significant differences between any of the comparisons. Although significant differences were not apparent, there was a borderline significant difference between Sham and TBI ($p = 0.0526$). The other comparisons (Sham vs. PMI, $p = 0.2846$, PMI larger; TBI vs. PMI, $p = 0.7338$ with TBI larger) were not significant. There were no interactions of note (interaction of day and group: $p = 0.3801$). As for days, day 7 was largest ($p < 0.001$ comparing to all others); consecutive days were not

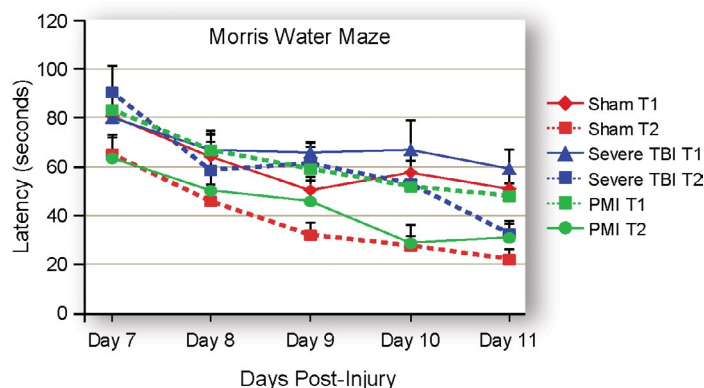


Figure 4: Effects of PMI-006 on the Morris Water Maze. The MWM is a test of hippocampal dependent spatial memory deficits. No significant differences were observed between any comparisons (Sham v. TBI + PMI-006, TBI v. TBI + PMI-006).

significantly different (8 vs. 9, $p=0.5324$; 9 vs. 10, $p=0.4935$; 10 vs. 11, $p=0.2572$), but nonconsecutive days were significantly different (8 vs. 10, $p=0.0142$; 9 vs. 11 $p=0.0028$). The effect was decreasing over time. There was also a significant effect of trial ($p<0.0001$).

Discussion

Despite decades of brain injury research and demonstrated success in pre-clinical studies using animal models of TBI, numerous clinical trials of neuroprotective agents have failed to show efficacy in human TBI patients [13,31]. Thus, we and others are still searching for novel therapeutic treatments that have the potential to mitigate TBI-induced neurodegeneration and improve functional outcome in TBI survivors. It is well established that injury-Induced inflammation and oxidative stress contribute significantly to deleterious secondary injury signaling cascades that result in progressive long-term neurodegeneration that is associated with lifelong disability in human TBI survivors [32,33]. Therapeutic treatments that affect these pathways could reduce neuronal death and improve functional outcome.

Here, we have described the molecular and functional effects of a new therapeutic drug candidate, PMI-006, a natural compound derived from the seeds of the *Aframomum melegueta* plant. Several studies have shown that extracts of *Aframomum melegueta* have strong anti-oxidant [34,35], anti-microbial [7,8], anti-apoptotic [6], anti-diabetic [36], anti-nociceptive [37] and anti-inflammatory [38] properties. Cumulatively, these properties suggest that this compound could have both analgesic and neuro protective effects after TBI. Our data is entirely consistent with these previous observations. The reduction of neuronal injury in the hippocampus correlates with the ameliorative effects of PMI treatment on TBI-induced inflammatory and oxidative stress signaling. Evidence suggests that drugs with pleiotropic properties, i.e., that possess both anti-inflammatory and anti-oxidative effects, appear to significantly improve functional outcome after TBI [39]. Because neuronal death from brain injury is due, in part, to a strong inflammatory response in the damaged brain tissue, these data support our hypothesis that natural product derived compounds with potent anti-inflammatory properties such as PMI 006 may reduce neuronal injury.

The demonstrated safety profile of *Aframomum melegueta* (e.g., its consumption by animals and humans) as well as its demonstrated protective effects in experimental models of human disease suggests that this compound is an excellent candidate for translational TBI studies. The potent neuroprotective properties of PMI may be due, in part, to its anti-inflammatory effects but PCR array data also show

that neuroprotection may be the result of its alteration of pro- and anti-oxidant pathways and its effects on NF κ B signaling (which is known to be activated after head injury) [40]. Although these studies were conducted in a rodent model of TBI, it has been shown that gene expression changes after TBI are commonly modulated across different species [41], suggesting that similar effects would be expected in human TBI patients. Natural compounds, such as PMI-006, that demonstrate such properties have great therapeutic potential for reducing neurodegeneration and improving functional outcome in TBI patients.

Acknowledgement

This study was supported by R21 NS053620, RO1 NS053620, The Moody Foundation, the University of Texas Medical Branch at Galveston Department of Anesthesiology, and the Medical student research program. We thank Christine Courteau-Butler and Andy Hall for excellent editorial support and Christy Perry for figures and illustrations. We thank Phytomedics, Inc. for freely and generously providing the *Aframomum melegueta* extract for our experimental brain injury studies.

References

1. Newman DJ, Cragg GM, Snader KM (2003) Natural products as sources of new drugs over the period 1981-2002. J Nat Prod 66: 1022-1037.
2. Newman DJ, Cragg GM (2012) Natural products as sources of new drugs over the 30 years from 1981 to 2010. J Nat Prod 75: 311-335.
3. Mishra B B, Tiwari V K (2011) Opportunity, Challenge and Scope of Natural Products in Medicinal Chemistry. Kerala India: 1-62.
4. Wilson RM, Danishefsky SJ (2006) Small molecule natural products in the discovery of therapeutic agents: the synthesis connection. J Org Chem 71: 8329-8351.
5. Lahlou M (2013) The success of natural products in drug discovery. Pharmacology & Pharmacy 4: 17-31.
6. El-Halawany AM, El Dine RS, El Sayed NS, Hattori M (2014) Protective effect of *Aframomum melegueta* phenolics against CCl₄-induced rat hepatocytes damage; role of apoptosis and pro-inflammatory cytokines inhibition. Sci Rep 4.
7. Konning GH, Agyare C, Ennison B (2004) Antimicrobial activity of some medicinal plants from Ghana. Fitoterapia 75: 65-67.
8. Ngwoke KG, Chevallier O, Wirkom VK, Stevenson P, Elliott CT, et al. (2014) *In vitro* bactericidal activity of diterpenoids isolated from *Aframomum melegueta* K.Schum against strains of *Escherichia coli*, *Listeria monocytogenes* and *Staphylococcus aureus*. J Ethnopharmacol 151: 1147-1154.
9. Hoge CW, McGurk D, Thomas JL, Cox AL, Engel CC, et al. (2008) Mild traumatic brain injury in U.S. Soldiers returning from Iraq. N Engl J Med 358: 453-463.
10. Shimamura M, Garcia JM, Prough DS, Hellmich HL (2004) Laser capture microdissection and analysis of amplified antisense RNA from distinct cell

- populations of the young and aged rat brain: effect of traumatic brain injury on hippocampal gene expression. *Mol Brain Res* 122: 47-61.
11. Hellmich HL, Eidson KA, Capra BA, Garcia JM, Boone DR, et al. (2006) Injured Fluoro-Jade-positive hippocampal neurons contain high levels of zinc after traumatic brain injury. *Brain Res* 1127: 119-126.
 12. Dopenberg EM, Choi SC, Bullock R (2004) Clinical trials in traumatic brain injury: lessons for the future. *J Neurosurg Anesthesiol* 16: 87-94.
 13. Xiong Y, Mahmood A, Chopp M (2009) Emerging treatments for traumatic brain injury. *Expert Opin Emerg Drugs* 14: 67-84.
 14. Beauchamp K, Mutlak H, Smith WR, Shohami E, Stahel PF (2008) Pharmacology of traumatic brain injury - where is the "golden bullet"?. *Mol Med* 14: 731-740.
 15. Langlois JA, Rutland-Brown W, Wald MM (2006) The epidemiology and impact of traumatic brain injury: a brief overview. *J Head Trauma Rehabil* 21: 375-378.
 16. Thornhill S, Teasdale GM, Murray GD, McEwen J, Roy CW, et al. (2000) Disability in young people and adults one year after head injury: prospective cohort study. *BMJ* 320: 1631-1635.
 17. Squire LR, Stark CE, Clark RE (2004) The medial temporal lobe. *Annu Rev Neurosci* 27: 279-306.
 18. Bast T (2007) Toward an integrative perspective on hippocampal function: from the rapid encoding of experience to adaptive behavior. *Rev Neurosci* 18: 253-281.
 19. Ginsberg SD, Hemby SE, Lee VM, Eberwine JH, Trojanowski JQ (2000) Expression profile of transcripts in Alzheimer's disease tangle-bearing CA1 neurons. *Ann Neurol* 48: 77-87.
 20. Morrison JH, Hof PR (2002) Selective vulnerability of corticocortical and hippocampal circuits in aging and Alzheimer's disease. *Prog Brain Res* 136: 467-486.
 21. Bayley PJ, Hopkins RO, Squire LR (2006) The fate of old memories after medial temporal lobe damage. *J Neurosci* 26: 13311-13317.
 22. Maguire EA, Burgess N, Donnett JG, Frackowiak RS, Frith CD, et al. (1998) Knowing where and getting there: a human navigation network. *Science* 280: 921-924.
 23. Maguire EA, Vargha-Khadem F, Mishkin M (2001) The effects of bilateral hippocampal damage on fMRI regional activations and interactions during memory retrieval. *Brain* 124: 1156-1170.
 24. Schmued LC, Albertson C, Slikker W Jr (1997) Fluoro-Jade: a novel fluorochrome for the sensitive and reliable histochemical localization of neuronal degeneration. *Brain Res* 751: 37-46.
 25. Hamm RJ (2001) Neurobehavioral assessment of outcome following traumatic brain injury in rats: an evaluation of selected measures. *J Neurotrauma* 18: 1207-1216.
 26. Ilic N, Schmidt BM, Poulev A, Raskin I (2010) Toxicological evaluation of grains of paradise (*Aframomum melegueta*) [Roscoe] K. Schum. *J Ethnopharmacol* 127: 352-356.
 27. Hellmich HL, Capra B, Eidson K, Garcia J, Kennedy D, et al. (2005) Dose-dependent neuronal injury after traumatic brain injury. *Brain Res* 1044: 144-154.
 28. (2004) SAS/STAT® 9.1 User's Guide. Cary, NC: SAS Institute.
 29. Smith C (2013) Review: the long-term consequences of microglial activation following acute traumatic brain injury. *Neuropathol Appl Neurobiol* 39: 35-44.
 30. Xiong Y, Shie FS, Zhang J, Lee CP, Ho YS (2004) The protective role of cellular glutathione peroxidase against trauma-induced mitochondrial dysfunction in the mouse brain. *J Stroke Cerebrovasc Dis* 13: 129-137.
 31. Schouten JW (2007) Neuroprotection in traumatic brain injury: a complex struggle against the biology of nature. *Curr Opin Crit Care* 13: 134-142.
 32. Ansari MA, Roberts KN, Scheff SW (2008) Oxidative stress and modification of synaptic proteins in hippocampus after traumatic brain injury. *Free Radic Biol Med* 45:443-452.
 33. Morganti-Kossmann MC, Rancan M, Otto VI, Stahel PF, Kossmann T (2001) Role of cerebral inflammation after traumatic brain injury: a revisited concept. *Shock* 16: 165-177.
 34. Onoja SO, Omeh YN, Ezeja MI, Chukwu MN (2014) Evaluation of the *In Vitro* and *In Vivo* Antioxidant Potentials of *Aframomum melegueta* Methanolic Seed Extract. *J Trop Med* 2014:1-6.
 35. Nwozo SO, Oyinloye BE (2011) Hepatoprotective effect of aqueous extract of *Aframomum melegueta* on ethanol-induced toxicity in rats. *Acta Biochim Pol* 58: 355-358.
 36. Adefegha SA, Oboh G (2012) Inhibition of key enzymes linked to type 2 diabetes and sodium nitroprusside-induced lipid peroxidation in rat pancreas by water extractable phytochemicals from some tropical spices. *Pharm Biol* 50: 857-865.
 37. Umukoro S, Ashorobi RB (2007) Further studies on the antinociceptive action of aqueous seed extract of *Aframomum melegueta*. *J Ethnopharmacol* 109: 501-504.
 38. Ilic NM, Dey M, Poulev AA, Logendra S, Kuhn PE, et al. (2014) Anti-inflammatory Activity of Grains of Paradise (*Aframomum melegueta* Schum) Extract. *J Agric Food Chem* 62: 10452-10457.
 39. Vink R, Nimmo AJ (2009) Multifunctional drugs for head injury. *Neurotherapeutics* 6: 28-42.
 40. Beni SM, Kohen R, Reiter RJ, Tan DX, Shohami E (2004) Melatonin-induced neuroprotection after closed head injury is associated with increased brain antioxidants and attenuated late-phase activation of NF-kappaB and AP-1. *FASEB J* 18: 149-151.
 41. Natale JE, Ahmed F, Cernak I, Stoica B, Faden AI (2003) Gene expression profile changes are commonly modulated across models and species after traumatic brain injury. *J Neurotrauma* 20: 907-927.