

QSAR Model for Androgen Receptor Antagonism - Data from CHO Cell Reporter Gene Assays

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Abstract

For the development of QSAR models for Androgen Receptor (AR) antagonism, a training set based on reporter gene data from Chinese hamster ovary (CHO) cells was constructed. The training set is composed of data from the literature as well as new data for 51 cardiovascular drugs screened for AR antagonism in our laboratory. The data set represents a wide range of chemical structures and various functions. Twelve percent of the screened drugs were AR antagonisms; three out of six statins showed AR antagonism, two showed cytotoxicity and one was negative. The newly identified AR antagonisms are: Lovastatin, Simvastatin, Mevastatin, Amiodaron, Docosahexaenoic acid and Dilazep.

A total of 874 (231 positive, 643 negative) chemicals constitute the training set for the model. The Case Ultra expert system was used to construct the QSAR model. The model was cross-validated (leave-groups-out) with a concordance of 78.4%, a specificity of 86.1% and a sensitivity of 57.9%. The model was run on a set of 51,240 EINECS chemicals, and 74% were within the domain of the model. Approximately 9.2% of the chemicals in domain of the model were predicted active for AR antagonism.

Case Ultra identified common alerts among different chemicals. By comparing biophores (alerts in positive chemicals) and biophobes (alerts in negative chemicals), it appears that chlorine (CI) and bromine (Br) enhance AR antagonistic effect whereas nitrogen (N) seems to decrease the effect. A specific study of benzophenones and benzophenone derivatives indicate that a radical with a "high" number of atoms in 4-position and/or other positions generally decrease the anti-androgenic effect.

Keywords: AR antagonism; CHO cells; Reporter gene assay; QSAR; Statins

Introduction

Inhibition of the Androgen Receptor (AR) dependent reporter gene transcription provides an important piece of information that flags potential endocrine-disrupting effect of a wide range of chemicals, e.g. pesticides, industrial chemicals and drugs [1-4]. *In vitro* data for AR antagonism may be used for priority setting for further studies, e.g. *in vivo* experiments that are more costly and time-consuming. By use of QSAR models for AR antagonism the priority capacity is enhanced considerably and is further enhanced when the QSAR models and associated training sets are improved. Pesticides and industrial chemicals dominate in our existing QSAR model for AR antagonism [5]. It is well known that some drugs have AR antagonistic effect either as a primary mechanism of action for efficacy or as a secondary mechanism not directly involved in the pharmacological action of the drug [6-8].

A primary antiandrogenic effect of bicalutamide and flutamide is used in treatment of prostate cancer [6,9,10]. Secondary anti-androgenic effect is found for the diureticum spironolactone and probably also for the antiarrhytmics quinidine, procainamide, disopyramide, sotalol, amiodarone, ibutilide and dofitilide [4,7,8,11-15]. Several epidemiological studies have related statins to improvement of prostate cancer treatment [16-19]. Statins are also related to improvement of treatment of acne and Polycystic ovary syndrome (PCOS) [20,21] - diseases that involve high testosterone levels. Drugs with antiandrogenic effect are known to be used against these diseases [6,22,23].

Spironolactone, quinidine, procainamide, disopyramide, sotalol, amiodarone, ibutilide, dofitilide and statins are well known Cardiovascular Disease (CVD) drugs [8,24]. An antiandrogenic effect of CVD drugs may thus be a possibility. Until now spironolactone is the only CVD drug present in the training set of our QSAR models for AR antagonism [4,5]. In this study our published QSAR model [5] was applied for analyzing the potential occurrence of AR antagonism among CVD drugs. 343 CVD drugs were screened. The QSAR model revealed biophores (chemical structures characteristic for active AR antagonism chemicals) in about 40% of the drugs. Chemical structures unknown to the QSAR AR antagonism model were also identified in 40% of the CVD drugs (data not published). Therefore it was decided to analyze some of the CVD drugs for AR antagonism in the AR reporter gene assay described previously [4]. The purpose of this study was to identify new AR antagonisms among the CVD drugs and to extend the domain of the future AR antagonism QSAR models. The selection of CVD drugs for AR reporter gene assay is described in material and methods.

Different QSAR models for AR antagonism have been published. Our QSAR model from 2008 was constructed by use of the software MultiCASE and 528 chemicals assayed by use of different cellular reporter gene assays [4]. Later, three modeling systems (MultiCASE, Leadscope and MDL QSAR) were used for the construction of AR antagonism models. There were 923-942 chemicals in the training sets also assayed by use of different cellular reporter gene assays [5].

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In addition, AR antagonism QSAR models based on only one single cell type and on a specific functional group, the brominated flame retardants (BFRs), have been published [25,26]. Recently, Kovarich et al. [27] used a training set consisting of AR antagonism data from 24 BFRs in a QSAR model developed especially for prediction of BFRs. Osteosarcoma cells from human (U2 OS) were used in the AR antagonism assay.

AR reporter gene assays are based on different cell types, e.g. Chinese hamster ovary (CHO-K1) cells [4], human mammary carcinoma cells (MDA-kb2) [28], U2 OS cells [29], African monkey kidney cells (CV-1) [30], human prostate adenocarcinoma PC-3 derived cell line (PALM) [31], human hepatoma liver cells (HepG2) [32] and yeast [33]. However, discrepancies between data from the different in vitro cell assays have been reported [34,35]. Previously it has been described that effectors which interact with the ligandreceptor complex, like response elements, corepressor or coactivator proteins, or other transcription factors, are cell-type dependent [36]. In addition, Kojima et al. [37] reported low sensitivity of reporter gene assays based on yeast cells, HepG2 cells or Hela cells (cervical cancer cells) as compared to reporter gene assays based on CHO cells. Thus, it may be an advantage to use data from only one single cell type for the development of AR antagonism QSAR models. Our database for AR antagonism contains data from 1140 chemicals, data from around 900 of these chemicals are based on CHO cells. As our AR antagonism data are collected continuously for all cell types, CHO cells are probably the most often used cell type for AR antagonism assays. Thus the aim of this study was to use AR antagonism data based exclusively on CHO cells to form a training set for a new "CHO" AR antagonism QSAR model. CHO data from an existing training set [5], new CHO data from the literature and new experimental data were used.

New software for QSAR modeling is developed continuously. In this study a newly developed program, Case Ultra, from MultiCASE Inc. was used. The program is especially suitable for the unbalanced training set [38] that was used in this study.

Materials and Methods

In vitro AR Assay

The AR antagonism assay was performed as previously described [4]. Shortly: Chinese hamster ovary cells (CHO-K1) were for each chemical run in two parallel lines; one transfected with the plasmids pSVAR0 and MMTV-LUC for antagonism, and another transfected with the plasmids pSVAR13 and MMTV-LUC for the cytotox evaluation. The MMTV-LUC plasmid contains gene coding for the reporter enzyme Luciferase. The plasmid pSVAR0 contains gene coding for the human androgen receptor (AR). The plasmid pSVAR13 contains gene coding for AR without a ligand-binding domain (LBD). CHO cells transfected with pSVAR0/MMTV-LUC need R1881 for AR activation. CHO cells transfected with pSVAR13/MMTV-LUC are constitutively AR activated. The chemicals were tested at concentrations of 1, 3, 10, and 30μ M, and within each assay all data were related to the response of 0.1 nM R1881 (methyltrienolone), which was set to 100%. IC_{25} , that is the concentration of test compound showing a 25% inhibition of the activity induced by 0.1 nM R1881, was calculated for each compound. The criteria for determining "a positive" was that a 25% inhibition of the 0.1 nM R1881-induced response should be reached at a non-cytotoxic concentration ≤10 µM. For QSAR modeling purpose, chemicals showing 25% inhibition at higher concentration than 10 µM belong to the group of "weak AR antagonisms/not AR antagonisms", also referred to as negatives.

Data for QSAR modeling

CHO data from literature: The data in the training set is comprised of experimental AR antagonism data from 15 publications [2-4,37,39-49]. The data represents a wide range of chemical structures and various functions and covers natural hormones, synthetic hormones and drugs, pesticides, Polyaromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), brominated flame retardants (BFRs), plasticizers and plastic additives, food additives and cosmetics, antioxidants, plant compounds, roast mutagens and various industrial chemicals. The data originates primarily from our laboratory of the National Food Institute, the Technical University of Denmark (laboratory 1) [4,42,45,48,49], the laboratory of Hokkaido Institute of Public Health, Japan (laboratory 2) [2,37,40,41,43] and the Endocrine Disrupting Chemical Analysis Center, Otsuka Life Science Initative, Otsuka Pharmaceutical Co. Ltd., 224-18 Ebisuno Hiraishi, Kawauchi-cho, Tokushima 771-0195, Japan (laboratory 3) [3,46]. Positive correlations were found between data from the various laboratories. Data reported $(IC_{20}(x) \text{ and } IC_{50}(y))$, respectively) from laboratory 2 and laboratory 3 found the equation: y = 3.589x + 0.3007 (R²= 0.6709, n=8), indicating an agreement with respect to potency between laboratories and a connection between IC-values usable as supplementary knowledge for grouping of the AR antagonism data.

All AR antagonism data were separated into two groups: a positive group (chemicals with $IC_{25} \le 10 \ \mu\text{M}$) and a negative group (chemicals with $IC_{35} > 10 \ \mu\text{M}$ or no activity).

Comparison of common data from the different laboratories showed a 83% (29/35)-91% (40/44) agreement. The compounds "2,4,5-trichlorophenoxyacetic acid" and "di-n-butyl phthalate" were indicated as weak AR antagonisms by Araki et al. [46] and this was in agreement with the finding in Vinggaard et al. [4], classifying the compound as negative. The remaining data was further evaluated. Some chemicals were excluded due to significant discrepancies between data without other supporting data (fenchlorphos, butyl benzyl phthalate, and the steroids estrone and corticosterone). Other data was excluded due to contradictory $IC_{_{25}}\!/IC_{_{20}}$ values close to 10 μM (fenvalerate, ethoxyquin). Laboratory 3 found IC $_{\rm 50}$ values of 26.9 and 35.9 μM for 4-tert-octylphenol and p-n-nonylphenol, respectively; according to laboratory 1, by using IC₂₅ as the cut-off, these two chemicals were classified "AR antagonism, high" and "negative", respectively. In the QSAR training set, 4-tert-octylphenol was set to positive and p-nnonylphenol to negative, also indicating that negative means either negative or weak positive. Takeuchi et al. [40] found the phthalate to be positive with an IC₂₀ value of 4.8 μ M. Due to more reports showing no AR antagonism, the di-n-butyl phthalate was included in the training set as a negative. In laboratory 3, dexamethasone was shown to have an IC_{50} value of 44.5 μ M, but was estimated by laboratory 1 to have an IC_{25} in the range of 1-3 μ M (AR antagonism, moderate); the decision was taken to include dexamethasone in the training set as a positive.

Benzophenones: Benzophenones represent a group of chemicals with functions as drugs and UV stabilizers in sunscreens, cosmetics and plastics [44]. Hydroxyl groups in benzophenones increase the antiandrogenic activity. The IC₅₀ value for benzophenone was 77 μ M and the calculated IC₂₅ was 29 μ M according to the reference [44]. Thus for the QSAR training set, benzophenone was classified as negative, while several of the hydroxylated benzophenone compounds were classified as positives [4,44].

Brominated flame retardants: BFRs are polybrominated diphenyl ethers and derivatives. In our previous QSAR model for AR antagonism

six BFRs analyzed by NFI were included in the training set [4,5]. New data from Kojima et al. [43] add further 15 BRFs to the training set. Analyzed by both laboratories (laboratory 1 and laboratory 2), BDE-100 was found to be positive [4,43].

Cardiovascular drugs: 343 cardiovascular drugs were identified in Drug References [8,24], and predictions were made for these compounds by our AR antagonism QSAR model [5]. The predicted activity (pos, neg, out of domain) as well as the presence of biophores, deactivating fragments and unknown chemical fragments was noted down. For the selection of CVD drugs for experimental testing in *in vitro* assay and inclusion of data in the training set, the following criteria for the drugs were used:

- Possible AR antagonism according to the literature [11-21]
- Part of a drug group
- The presence of a biophore
- The presence of an unknown fragment
- Overall a distribution between positive and negative corresponding to about 25% and 75%, respectively; two times the presence of positives previously found for the EINECS chemicals [4,5].

100 drugs were selected due to these criteria. 51 of the drugs were directly commercially available and characterized as: 26 out of domain (QSAR predicted), 25 within domain with 7 positives and 18 negatives; 40% of the drugs contained biophores and 40% contained unknown chemical fragments. Separated into pharmacological groups, the distribution was as follows, with the number given in parenthesis: Alpha blockers (1)*, angiotensin II receptor antagonisms (5), antiarrhythmics (17), lipid-regulating statins (6), lipid-regulating fibrates (5), lipid-regulating nicotinates (1), lipid-regulating bile acid-binding resins (1), triglyceride-reducing polyunsaturated fatty acids (2), direct-acting vasodilators, (4), vasodilators for ischaemic heart disease (1), and Vasodilators for cerebral and peripheral vascular disorders (7).

*also belong to the direct-acting vasodilators.

The whole process for the 51 selected CVD drugs, from QSAR prediction to *in vitro* laboratory experiments to the prediction by our new QSAR model, is described in Supplement 1. Supplement 1 is available at http://qsar.food.dtu.dk/AntiAndrogensup1.zip

Data preparation for QSAR modeling: For the QSAR modeling the chemical structures were described using SMILES (simplified molecular input entry system) and imported into OASIS DataBase Manager (DBM) [50]. In DBM, a hydrolysis simulation was performed and examination for chemicals without at least two carbons (including inorganics) and chemicals containing heavy atoms. Salts (e.g. sodium/ potassium-salts and hydrochlorides) were analyzed and the ones not containing toxic ions were processed by removing the ion part(s) from the structure. Duplicate or conflicting occurrences were removed from the structure set. Thereafter some SMILES codes were removed due to the MultiCase procedure for checking of SMILES and Data Kurator, the Case Ultra procedure for additional checking of SMILES for correctness.

During creation of the model, α hexachlorocyclohexane and β hexachlorocyclohexane were identified to have the same 2-D-structure (identical SMILES) and also having the same activity; only one was included in the training set. Dieldrin and Endrin were identified to having the same structure but different activities; none of them were included in the training set.

The training set is available as supplement 2 at http://qsar.food. dtu.dk/AntiAndrogensup 2.zip, and contains information on CAS numbers, machine-readable structure notations and activities.

Modeling methodology

Algorithm: The Case Ultra 64-bit 1.4.0.0 modeling system from Multicase Inc. was used [38]. It is a further development of MC4PC, MultiCASE, previously used [4,5]. Case Ultra uses SMILES codes to enter chemicals. The program is a fragment (alert)-based statistical model system that aims to discover fragment combinations, which are relevant for the observed effect. Biophores are structural alerts that appear mostly in active molecules and therefore may be responsible for the observed activity. Case Ultra also looks at inactivities in the training set to identify deactivating fragments, deemed biophobes. Case Ultra has new functionalities and features and a new algorithm to discover structural alerts. New descriptors are added, e.g. estate values, surface and volume descriptors, Gasteiger atom based charges, vapor pressure, pKa and hydrogen bond donor/acceptors. Alerts are no longer only linear or with only one branch. They are now more general substructures. More model validation options exist, e.g. leave N% out N times, also for unbalanced training sets [38]. Case Ultra also uses physicochemical data (e.g. log (octanol/water) partition coefficient) as well as pharmacokinetic data (e.g. Lipinski rule of five and human intestinal absorption) and fragments as modulators (increasing or decreasing the activity prediction).

Applicability domain: While making a prediction, Case Ultra may report that the prediction is out of domain; this may be due to the presence of fragments not occurring in the training set. In the Case Ultra domain definition, up to one unknown fragment is accepted.

Predictions may also be inconclusive, e.g. when a chemical contains biophores as well as biophobes.

Statistical analysis: In Case Ultra a specific program for unbalanced training sets is available and used in this study. Leavegroups-out cross-validation was used. The Case Ultra model was validated five times two-fold 50% cross-validation [38,51]. The crossvalidation result was evaluated by use of Cooper statistics [52]. Cooper statistics use sensitivity (ability to predict actives), specificity (ability to predict inactives), and concordance (overall accuracy) to describe the predictivity of a model.

Results

Cardiovascular drugs in the training set

Data of the newly assayed cardiovascular drugs are shown in table 1. AR antagonism for drugs in concentration $\leq 10 \ \mu$ M was found in six drugs out of 51. This corresponds to AR antagonism in 12% of the investigated cardiovascular drugs.

The initial QSAR prediction with our published MultiCase model [5] of 17 antiarrhytmics showed defitilide and ibutilide to be positive due to the presence of biophores. However, the molecules also contain deactivating fragments (biophobes). Only amiodarone among the antiarrhytmics contains solely a biophore. The *in vitro* assay showed that only amiodarone among the antiarrhytmics was positive. Among the lipid-regulating CVD drugs, two statins (lovastatin and simvastatin) out of six were predicted to be positive due to biophores only. These two statins were also found to be positive in the laboratory test. In addition, mevastatin was experimentally found to be positive. The QSAR prediction of atorvastatin, fluvastatin and pravastatin showed all three to contain biophores as well as unknown fragments. The *in*

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vitro assay showed atorvastatin and fluvastatin to be cytotoxic, while pravastatin was negative (Figure 1).

Among five lipid-regulating fibrates, only negative *in vitro* results were found. This was partly in agreement with the QSAR predictions

Drug group	Drug name	CAS no. for QSAR analysis	CAS no. for experimental assay	Classification
Alpha blockers	Tolazoline*	59-98-3	59-97-2	neg
Angiotensin II receptor antagonisms	Irbesartan	138402-11-6	a	neg
	Losartan	114798-26-4	a	neg
	Valsartan	137862-53-4	a	neg
	Telmisartan	144701-48-4	a	neg
	Eprosartan	133040-01-4	a	neg
Antiarrhythmics	Amiodaron	1951-25-3	a	AR antagonism, low
	Disopyramid	3737-09-5	a	neg
	Propafenon	54063-53-5	34183-22-7	neg
	Sotalol	3930-20-9	959-24-0	neg
	Flecainid	54143-55-4	54143-56-5	neg
	Lidocain	137-58-6	a	neg
	Mexiletin	31828-71-4	a	neg
	Phenytoin	57-41-0	a	neg
	Verapamil	52-53-9	152-11-4	tox
	Hydroquinidine	1435-55-8	a	neg
	Procainamide	51-06-9	614-39-1	neg
	Quinidine	56-54-2	a	neg
	Tocainide	41708-72-9	71395-14-7	neg
	Bretylium	59-41-6	61-75-6	neg
	Acecainide	32795-44-1	a	neg
	Dofetilide	115256-11-6	a	neg
	Ibutilide	122647-31-8	122647-32-9	neg
Lipid-regulating, statins	Atorvastatin	134523-00-5	134523-03-8	tox
	Fluvastatin	93957-54-1	93957-55-2	tox
	Lovastatin	75330-75-5	a	AR antagonism, high
	Pravastatin	81093-37-0	81131-70-6	neg
	Simvastatin	79902-63-9	а	AR antagonism, high
	Mevastatin	73573-88-3	a	AR antagonism, moderate
Lipid-regulating, fibrates	Benzafibrat	41859-67-0	а	neg
	Gemfibrozil	25812-30-0	а	neg
	Ciprofibrate	52214-84-3	а	neg
	Clofibrate	637-07-0	а	neg
	Fenofibrate	49562-28-9	а	neg
Lipid-regulating, nicotinates	Acipimox	51037-30-0	а	neg
Lipid-regulating, bile acid-binding resins	Colestipo	37296-80-3	а	neg
Triglyceridereducing, Polyunsaturated	Docosahexaenoic acid	6217-54-5	а	AR antagonism, low
fatty acids	Eicisapentaenoic acid	10417-94-4	a	neg
Vasodilators, direct-acting	Diazoxide	364-98-7	a	neg
	Hydralazine	86-54-4	304-20-1	neg
	Minoxidil	38304-91-5	a	neg
	Todralazine	3778-76-5	a	neg
Vasodilators, ischaemic heart disease	Dilazep	35898-87-4	a	AR antagonism, moderate
Vasodilators, cerebral, peripheral vascular	Buflomedil	55837-25-7	35543-24-9	neg
disorders	Cyclandelate	456-59-7	а	neg
	Fasudil	103745-39-7	203911-27-7	neg
	lfenprodil	23210-56-2	a	neg
	Inositol nicotinate	6556-11-2	a	neg
	Nicotinyl alcohol	100-55-0	a	neg
	Pentifylline	1028-33-7	a	neg
	Pentoxifylline	6493-05-6	a	neg

*also a Vasodilator, direct acting

Table 1: Cardiovascular drugs: The name and CAS number of each drug as well as the classification of the drug as positive (AR antagonism), negative or toxic are shown. The drugs were tested at 1, 3, 10, and 30 μ M. Potency codes for positive drugs were as follows: high, 0.3 μ M < IC₂₅ ≤ 1 μ M; moderate, 1 μ M < IC₂₅ ≤ 3 μ M; low, 3 μ M < IC₂₅ ≤ 10 μ M; neg, negative chemicals; tox, cytotoxic chemicals. Non-cytotoxic chemicals identified "low – high" were classified as positive for AR antagonism. For some drugs, two CAS nos. are given, one for QSAR use and one for the experimental assay, i.e. only non-salts versions of drugs are used for QSAR modeling. ^aCAS no. identical with CAS no. for QSAR analysis.



showing four out of the five drugs to be negative. Fenofibrate was predicted positive. QSAR prediction of two other lipid-regulating CVD drugs, acipimox and colestipol, showed acipimox to contain unknown fragments and colestipol to be negative. The *in vitro* assay showed both to be negative.

Among the polyunsaturated fatty acids, the *in vitro* assay showed docosahexaenoic acid to be positive.

For two of the CVD drug groups, the "Angiotensin II receptor antagonisms" and the "Vasodilators, direct acting", the QSAR prediction showed all (five and five, respectively) to be out of domain due to the presence of unknown chemical fragments. Four of the "Angiotensin II receptor antagonisms" also contained biophores. The *in vitro* assay showed all to be negative. Among the other vasodilators, the vasodilator used for ischaemic heart disease (dilazep) was QSAR predicted to contain solely biophores and was found to be positive in the *in vitro* assay.

Lovastatin, simvastatin, mevastatin, amiodaron, docosahexaenoic acid and dilazep were included in the QSAR training set as positives. The other drugs were included as negatives, except for the chemicals producing cytotoxicity (two statins, atorvastatin and fluvastatin, and one antiarrhythmic, verapamil), which were excluded from the training set.

Data for 890 chemicals from the existing training set, new CHO data from the literature and the new experimental data for the CVD drugs made up for the new training set. These chemicals were reduced to 874 chemicals. Among the newly analyzed CVD drugs, acipimox and bretylium were not accepted during the technical adaptation procedure.

Validation, applicability domain and chemicals with antiandrogenic effect

The five times two-fold 50% cross-validation of the Case Ultra QSAR model (Table 2) showed a sensitivity of 57.9%, a specificity of 86.1% and a concordance of 78.4%.

In total, 51,240 discrete organic EINECS chemicals (European INventory of Existing Commercial chemical Substances) were predicted using the modeling system. Table 2 shows the domain of the Case Ultra model to be 74% of the EINECS chemicals. The percentage of the screened EINECS chemicals that were predicted positive for AR antagonism was 9.2%.

Biophores

From the 874 chemicals in the training set for the Case Ultra model, 79 alerts were identified, 45 as biophores and 34 as biophobes. 35 of the biophores were present in three or more active molecules in the training set. Table 3 shows the most significant alerts in the training set of the Case Ultra model; in addition, other alerts were shown. The most significant alerts (Alert no. 6 (cc(ccc)c) and alert no. 1 ((Cl)ccccc)) were primarily found in PCBs. Other alerts, e.g. no. 4 (c1ccccc1), were found in PCBs and chemicals from other groups (e.g. brominated diphenyl ethers) as well.

Biophore no. 15 (c(O)cccc(O)) was found solely in benzophenones. Benzophenones with the hydroxyl group in 2-position contain biophore no. 15. Another biophore (no. 5, (c(c(cc)=O)C)) was found in other benzophenones as well as in other chemicals. Although experimentally negative benzophenone contains alert no. 5 (17 out of 23 molecules containing this alert are active). In addition, benzophenone also contains a deactivating fragment (alert no. 75(c1(C(c2cccc2))ccccc1, 0 out of 2 molecules containing this alert are active). The negative hydroxylated

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Software Chemicals accepted by the modeling system		Chemicals in domain*	Chemicals in domain with AR antagonism activity		
	Number (%)	Number (%)	Number (%)		
Case Ultra	45,511 (100)	33,715 (74)	3,090 (9.2)		

*chemicals predicted marginal or inconclusive are not included

 Table 2: The CHO AR antagonism model applied to 51,240 EINECS chemicals.

Biophores		Example of a molecule containing the alert and main chemical group		Statistical significance ^c	Molecu	Molecules containing alerts		
no	Structural alert ^a	Chemical name/ main chemical group	CAS no.	%	total	active	inactive	
6	cc(ccc)c	2,3',4,4',5-Pentachlorobiphenyl/ Biphenyls	31508-00-6	100	18	17	1	
1	(CI)ccccc	Dichlofenthion/ polychlorbiphenyls	97-17-6	99.8	35	26	9	
15	c(O)cccc(O)	2,4-Dihydroxybenzophenone/ Benzophenones	131-56-6	99.6	8	8	0	
2	c(Clccc)ccCl ^b	2,2',3',4,4',5-Hexachlorobiphenyl/ polychlorbiphenyls	35065-28-2	99.7	26	20	6	
3	(Br)ccc	O-(4-Bromo-2,5-dichlorophenyl) O-methyl phenylphosphonothioate/ Brominated diphenyl ether	21609-90-5	99.5	25	19	6	
4	c1ccccc1 ^b	2,3,3',4,4'-Pentachlorobiphenyl/ Brominated diphenyl ether, polychlorbiphenyls	32598-14-4	99.5	25	19	6	
7	c(cc)cc	2,3',4,4'-Tetrachlorobiphenyl/ Biphenyls	32598-10-0	99.5	20	16	4	
5	Ccc(C(=O)cc)cc ^b	Chlorohydroxy benzophenone/ benzophenones	85-19-8	98.8	23	17	6	
8	Cc1ccccc1	4-(1,1,3,3-Tetramethylbutyl)phenol/ methylbenzene	140-66-9	97.8	19	14	5	
10	cc(OP(=S)O)	Dichlofenthion/Phosphorbenzene	97-17-6	96.7	18	13	5	
11	(CI)ccc(CI)	2,4,6-Trichlorophenylhydrazine/ chlorbenzene	5329-12-4	96.7	18	13	5	
12	ccc(cc)c	7,12-Dimethylbenz(a)anthracene/ PAHs	57-97-6	95.8	15	11	4	
13	cccc(O)c	Amiodaron/no main chemical group, structure: oxygen linked to ring structure	1951-25-3	94.6	12	9	3	
27	*	Mevastatin	73573-88-3	87.5	3	3	0	
*c(=o)occ2c(=c)ccc2CCc1cc	(O)cc(=O)o1						
26	ccc(c(C))	Mevastatin	73573-88-3	84.4	5	4	1	
33	CC(C)C	Simvastatin	79902-63-9	75.0	4	3	1	

^astructural alerts are described using SMILES notation for aliphatic and aromatic compounds according to the Daylight Theory Manual [59]. Notation: Lower case atoms are aromatic.

(): branch point. When the alert covers part of an aromatic structure, the attached part of the alert (aliphatic or aromatic) is enclosed in parenthesis. ^btwo occurrences of the same biophore in the mentioned molecule.

°(1-p-value) x 100

Table 3: The most significant alerts (biophores) and the alerts in the active CVD drugs in the Case Ultra AR antagonism model.

benzophenone derivatives containing alert no. 5 were characterized by having a radical with a "high" number of atoms in 4-position and/ or other positions, e.g. octyloxy, sulfonic acid, dibutylamino and 2-methylpropanoic acid 1-methylester. The CVD drug fenofibrate belongs to this group of negative benzophenone derivatives and also contains a deactivating fragment (alert no. 64 (Cc1ccccc1O) 0 out of 7 molecules containing this alert are active) (Figure 2).

Alert no. 12 (ccc(cc)c) was mostly found in PAHs with a distribution between positive and negative of 75%.

Alert no. 13 (cccc(O)c) was present in two of the new identified AR antagonisms with cardiovascular drug effect (amiodarone - an antiarrythmics, and dilazep - a vasodilator used for treatment of ischaemic heart disease). Other chemicals with this alert were herbicides, fungicides and BRFs. Figure 3 shows alert no. 13 in prochloraz (a fungicide), BDE-100 (a BFR) and amiodarone (a CVD drug). Distribution between positive and negative chemicals in this alert group was 75%.

The three active statins contained a common alert (no. 27), an alert not found in other types of chemicals; the alert contained a high number of carbons (16) and oxygen (5). However, as in the initiating MultiCASE QSAR prediction, lovastatin and simvastatin differ from mevastatin; in addition, lovastatin and simvastatin contained biophore

no. 33 also found in 3-methylpent-1-ene and 1-hexene,3 methyl. Mevastatin also contained an additional alert (no. 26), an alert which was common with spironolactone, betulin, corticosterone acetate and mifepristone, all steroid-like drugs. For the last new drug with AR antagonistic effect "Docosahexaenoic acid", the alert was the whole molecule.

Deactivating fragments (Biophobes)

Table 4 shows the alerts in the inactive chemicals (the biophobes). Nitrogen is present in four out of ten most significant biophobes, two in ring structures. Sulfur and phosphorus were present in one alert each; otherwise carbon atoms and oxygen make up the alerts. The most significant biophobe was alert no. 46 consisting of a nitrogencarbon structure ([n]c) placed in ring structures, 57 out of 58 molecules containing [n]c are inactive. Among the CVD drugs, all five Angiotensin II receptor antagonisms contain alert no. 46. In addition, three other of the CVD drugs also contained alert no. 46. Alert no. 47 consists of the hydroxylated part (CO) of organic acids, 51 out of 54 molecules containing (CO) are inactive. Many of the inactive cardiovascular drugs contribute to the training set with molecules containing alert no. 47. However, the two mentioned polyunsaturated fatty acids experimentally found to be AR antagonists (docosahexaenoic acid and γ -linolenic acid) also contain alert no. 47.

Biophobes		Example of a molecule containing the alert ar structures	Statistical significance ^b	Molecules containing alerts			
no	Structural alert ^a	Chemical name/ main chemical structure	CAS no.	%	total	active	inactive
46	[n]c	Hydralazine/ nitrogen containing ring structure	86-54-4	100	58	1	57
47	со	Pravastatin/ hydroxylated compounds in acids	81093-37-0	100	54	3	51
48	cc(=O)c	Benzanthrone/unsaturated oxygen linked to ring structure	82-05-3	100	35	0	35
49	S(=0)=0	Sulfanilamide/ sulfon compounds	63-74-1	100	37	2	35
50	CCCCCC	Hexyl cinnamic aldehyde/ lineaer saturated carbon structure	101-86-0	100	34	2	32
51	ccn	Quinoline/ nitrogen in ring structure	91-22-5	100	26	1	25
52	(c(O)cc)o	Quercetin/hydroxylated compounds linked to benzene	117-39-5	100	26	2	24
53	C=N, c=n	Tolazoline/unsaturated nitrogen, carbon in aromatic as well as aliphatic structure	59-98-3	100	23	1	22
54	cc(cC)cc	p-Toluenesulfonamide	70-55-3	100	23	2	21
55	P=O	Trichlorofon/unsaturated phosphor, oxygen linear structure	52-68-6	100	19	0	0
56	ccc(N)cc	1-Naphthylamine, N-phenyl-/ N-phenyl	90-30-2	100	19	2	17
59	c(C(=O)N)	Flecainide	54143-55-4	100	13	0	13
61	cCc	Ifenprodil	23210-56-2	99.7	12	1	11
64	Cc1ccccc1O	Fenofibrate	49562-28-9	99.2	7	0	7
75	c1(C(c2cccc2))ccccc1	Benzophenone	119-61-9	75	2	0	2

^astructural alerts are described using SMILES, see footnote to Table 3. ^b(1-p-value) x 100

Table 4: The most significant alerts (biophobes) in the inactive molecules and the alerts in the inactive CVD drugs and benzophenone in the Case Ultra AR antagonism model.

Discussion

The CVD drugs and the AR antagonism

The in vitro assay showed an occurrence on 12% of AR antagonisms among cardiovascular drugs. This is at the level previously found among 49,292 EINECS chemicals [5]. However, as for the EINECS chemicals, some drug groups contain AR antagonisms more frequently than others. Among the antiarrhytmics only amiodarone was positive. The presence of antiandrogenic effect among statins was confirmed. An intuitive evaluation of figure 1 indicates that the AR antagonism activity of statins may be related to the pyran part and the cytotoxicity to the azole, benzene fluoride part. The presence of antiandrogenic effects of statins and not of fibrates and acipimox and colestipol are supported by the findings in a Finnish epidemiological study. This study showed statins to have a beneficial effect on prostate cancer prevention; this effect was not found for fibrates, acipimox and colestipol [19]. The finding of docosahexaenoic acid as positive was unexpected, but another polyunsaturated fatty acid "y-linolenic acid" was also found to be positive in a previous study [4]. The vasodilator used for ischaemic heart disease (dilazep) was also found to be positive. This in vitro finding was expected only due to the prediction by our published QSAR model [5]. This study shows that CVD drugs that were predicted to be positive by our previous published QSAR model were generally confirmed to be positive in the *in vitro* assay.

The new QSAR model

The concordance of 78.4% was slightly higher than the concordances of our previous QSAR models for AR antagonism [4,5]. Modeling the newly developed training set by use of the MultiCASE software, which we have used for previous QSAR modeling [5], a similar concordance on 78.7% was found. The domain of the present Multicase Ultra model was improved from about 60% to 74% as compared to our previous AR antagonist QSAR models [4,5].

Alerts (biophores/biophobes) and the CVD drugs

In some cases, an individual alert can make up the main part of a chemical group, e.g. PCBs or benzophenones, but can also be common for chemicals belonging to different chemical groups. PCBs which in particular are AR antagonists (74% of 39 PCBs was experimentally shown to be AR antagonists [4]) does not have common biophores with the CVD drugs in this study.

Evaluating positive and negative chemicals in the alert groups it was found that a radical with a "high" number of atoms in 4-position or/and other positions may decrease the AR antagonism of benzophenones. The CVD drug fenofibrate contains the actual biophore but also a radical with a "high" number of atoms in 4-position and a significant biophobe and perform no AR antagonism. This finding adds new knowledge to the identification of chemical structures of importance for the AR antagonism of Benzophenones. Kowamura et al. [44], has previously shown that a hydroxylated group at the 2-position generally enhances the antiandrogenic activity of benzophenones (Figure 2). Thus evaluating positive and negative chemicals in the alert groups of QSAR models may be a useable supplementary tool for potency evaluation.

Alert group no. 13 (cccc(O)c) was present in chemical groups with different functions, e.g. prochloraz (a fungicide), BDE-100 (a BFR) and amiodarone/dilazep (CVD drugs). This illustrates that it is valuable to include a wide range of chemical structures in the training set (Figure 3).

The three statins performing AR antagonism were present in one alert group (no. 27). This biophore contained the pyran part (Figure 1). Lovastatin/simvastatin and mevastatin were also included in an additional biophore group, respectively. Mevastatin was in the same biophore group as steroid-like drugs, e.g. spironolactone. The statin with no AR antagonism "Pravastatin" was in the large alert group



Figure 2: The AR antagonism of Benzophenone and Benzophenone derivatives. The effect of hydroxylation and radical substitution is illustrated. (A) Benzophenone - a weak AR antagonism, (B) 4-hydroxy-4'-chlorobenzophenone - a strong AR antagonism, (C) Fenofibrate - no AR antagonism.



Figure 3: A common biophore (alert no. 13 (cccc(O) c) in chemicals with different primary functions. (A) a fungicide (Prochloraz, CAS no. 67747-09-5), (B) a brominated flame retardant (BDE-100, CAS no. 189084-64-8) and (C) a drug (Amiodarone, CAS no. 1951-25- 3), respectively. The common biophore is marked.

no. 47 (CO), strongly indicating no AR antagonism. Using our new QSAR model on the experimentally cytotoxic statins fluvastatin and atorvastatin, the prediction showed both to be "inconclusive" due to content of biophores as well of biophobes. Acipimox and bretylium,

not accepted due to the technical procedure, were predicted to be out of domain.

Case Ultra makes use of the ratio between active/inactive when predictions are made and chemicals like fatty acids with unexpectedly AR antagonistic activity would probably be predicted as inactive AR antagonisms, not least due to the biophobe no. 47. Only one biophore will exist for such chemicals and cover the whole molecule. Thus, in spite of having QSAR models for important priority setting, experimental measurement should be performed when possible. The AR antagonism of docosahexaenoic acid and γ -linolenic acid [4] is a good example of this. This property of these unsaturated fatty acids deserves further attention.

By comparing biophores and biophobes it appears that chlorine (Cl) and bromine (Br) enhance the AR antagonistic effect while nitrogen seems to decrease the effect.

Comparison of AR antagonism data between assays based on different cell types

In general, a good agreement exists between qualitative data from various AR reporter gene assays, and QSAR models have been developed by use of data from different assays [4,5]. Minor disagreements between data for AR antagonism in CHO cells and U2-OS cells are found [4,29,43]. A comparison of AR antagonism data for brominated flame retardants experimentally performed on U2-OS cells as well as on CHO cells shows a possible disagreement on 27% (3/11), i.e. differences were found for BDE-153, BDE-190 and TBBPA.

Disagreement between AR antagonism in CHO cells and MDA (human mammary carcinoma) cells was also found; i.e. chemicals with AR antagonism in CHO cells were inactive in MDA cells with steroids dominating, making up three (progesterone, cyproterone acetate and 17 β -estradiol) out of four chemicals [4,28,53]. The fourth chemical was chlordane [2,34,54]. This disagreement for chlordane was also reported previously by Aït-Aïssa et al. [34]. For chemicals showing AR antagonism in MDA cells but inactive in CHO cells, all were non-steroids (phenanthrene, pirimiphosmethyl, chlorpropham, metolachlor, pretilachlor) [2,4,28,34,55]. Chemicals like diethyl phthalate, aldrin and γ -lindane having IC₅₀ values at concentrations of 60 - 80 μ M in MDA cells were negative in CHO cells [4,28,40] where the cut-off for positive results was set at 10 μ M. However, due to the relatively high concentration needed for AR antagonism in MDA cells, these chemicals were considered negative in the two cell assays.

Thus, discrepancies in AR antagonism between cellular assays should without a doubt be taken into account when QSAR models are to be developed. Data from some cell types seem more appropriate to include in the same training set than others, without exclusion of too many contradicting results. Individual cellular QSAR models as well as multicellular QSAR models for AR antagonism could be developed with advantage.

Discrepancies in AR antagonism between cells may be due to difference in mechanism as well as the used cell type. Differences may be: initiating ligands (R1881, DHT) for the AR, metabolism, reporter plasmid, cellular presence of endogeneous receptors (AR, glucocorticoid receptor (GR)), cytotoxicity (transcription complex level (used in CHO assays), use of phase-contrast microscopy for cellular vacuolization (in MDA assays), etc.) [2,4,28,55].

In addition, discrepancies in AR antagonism between cells may be due to chemical specific capacity for agonism and Luciferase

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inhibition [3,45,56-58]. AR agonists are very rare among industrial and environmental chemicals (2/502 =0.5%) [2,3,39,43,44,47], but may be rather related to the steroid structure [3,35].

Deviations between data from different CHO assays may also occur. This may be related to minor differences between protocols. Thus some laboratories use R1881 to initiate the AR mediated transcription, other laboratories use DHT. Some laboratories report IC₂₅ and measure inhibition up to 10 μ M; other laboratories report IC₅₀ and measure inhibition up to 100 μ M or higher. Compared to other cellular AR antagonism protocols, the applied assays in the CHO assays for cytotoxicity seem relatively similar, i.e. the cytotoxicity of chemicals is evaluated by transcriptional activity [2,4], which is the ideal way to investigate cytotoxicity in this context.

Conclusion

In the development of QSAR models for Androgen Receptor (AR) antagonism, a training set based on Chinese hamster ovary (CHO) cells was constructed. Data from the literature and data on 51 cardiovascular drugs recently screened for AR antagonism at our laboratory at the National Food Institute (NFI) make up the training set. All data together represent a wide range of chemical structures and various functions. Twelve percent of the NFI screened drugs are AR antagonisms; 3 out of 6 statins showed AR antagonism, two showed cytotoxicity and one was negative. Newly identified AR antagonisms are: Lovastatin, Simvastatin, Mevastatin, Amiodarone, Docosahexaenoic acid and Dilazep.

A total of 874 (231 positive, 643 negative) chemicals constitute the training set for the model. The Case Ultra expert system was used to construct the QSAR model. Case Ultra showed a concordance of 78.4%, a sensitivity of 57.9% and a specificity of 86.1%. The model was run on a set of 51,240 EINECS chemicals, and 74% were within the domain of the model. Approximately 9.2% of the chemicals in the model domain were predicted active for AR antagonism.

Case Ultra identifies common alerts among chemical groups with equal as well different functions. By comparing biophores and biophobes, it appears that Cl and Br may enhance AR antagonistic effect while nitrogen seems to decrease the effect. A specific study of benzophenones and benzophenone derivatives indicate that a "high" number of atoms in 4-position and/or other positions generally decrease the antiandrogenic effect.

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References

- Jensen GE, Niemelä JR, Wedebye EB, Nikolov NG (2008) QSAR models for reproductive toxicity and endocrine disruption in regulatory use--a preliminary investigation. SAR QSAR Environ Res 19: 631-641.
- Kojima H, Katsura E, Takeuchi S, Niiyama K, Kobayashi K (2004) Screening for estrogen and androgen receptor activities in 200 pesticides by in vitro reporter gene assays using Chinese hamster ovary cells. Environ Health Perspect 112: 524-531.
- Araki N, Ohno K, Nakai M, Takeyoshi M, lida M (2005) Screening for androgen receptor activities in 253 industrial chemicals by in vitro reporter gene assays using AR-EcoScreen cells. Toxicol In Vitro 19: 831-842.

- Vinggaard AM, Niemelä J, Wedebye EB, Jensen GE (2008) Screening of 397 chemicals and development of a quantitative structure--activity relationship model for androgen receptor antagonism. Chem Res Toxicol 21: 813-823.
- Jensen GE, Nikolov NG, Wedebye EB, Ringsted T, Niemela JR (2011) QSAR models for anti-androgenic effect--a preliminary study. SAR QSAR Environ Res 22: 35-49.
- Reid P, Kantoff P, Oh W (1999) Antiandrogens in prostate cancer. Invest New Drugs 17: 271-284.
- Berardesca E, Gabba P, Ucci G, Borroni G, Rabbiosi G (1988) Topical spironolactone inhibits dihydrotestosterone receptors in human sebaceous glands: an autoradiographic study in subjects with acne vulgaris. Int J Tissue React 10: 115-119.
- Sweetman SC (2005) Dose adjustment in renal impairment: response from Martindale: the Complete Drug Reference. BMJ 331: 292-293.
- Gravina GL, Festuccia C, Galatioto GP, Muzi P, Angelucci A, et al. (2007) Surgical and biologic outcomes after neoadjuvant bicalutamide treatment in prostate cancer. Urology 70: 728-733.
- Nishimura K, Arichi N, Tokugawa S, Yoshioka I, Kishikawa H, et al. (2007) Effects of flutamide as a second-line agent for maximum androgen blockade of hormone refractory prostate cancer. Int J Urol 14: 264-267.
- Rose LI, Underwood RH, Newmark SR, Kisch ES, Williams GH (1977) Pathophysiology of spironolactone-induced gynecomastia. Ann Intern Med 87: 398-403.
- Makkar RR, Fromm BS, Steinman RT, Meissner MD, Lehmann MH (1993) Female gender as a risk factor for torsades de pointes associated with cardiovascular drugs. JAMA 270: 2590-2597.
- Gowda RM, Khan IA, Panukollu G, Vasavada BC, Sacchi TJ, et al. (2004) Female preponderance in ibutilide-induced torsade de pointes. Int J Cadiol 95: 219-222.
- Fülöp L, Bányász T, Szabó G, Tóth IB, Bíró T, et al. (2006) Effects of sex hormones on ECG parameters and expression of cardiac ion channels in dogs. Acta Physiol (Oxf) 188: 163-171.
- Pham TV, Sosunov EA, Anyukhovsky EP, Danilo P Jr, Rosen MR (2002) Testosterone diminishes the proarrhythmic effects of dofetilide in normal female rabbits. Circulation 106: 2132-2136.
- Platz EA, Leitzmann MF, Visvanathan K, Rimm EB, Stampfer MJ, et al. (2006) Statin drugs and risk of advanced prostate cancer. J Natl Cancer Inst 98: 1819-1825.
- Murtola TJ, Visakorpi T, Lahtela J, Syvälä H, Tammela TLj (2008) Statins and prostate cancer prevention: where we are now, and future directions. Nat Clin Pract Urol 5: 376-387.
- Friedman GD, Flick ED, Udaltsova N, Chan J, Quesenberry CP Jr., et al. (2008) Screening statins for possible carcinogenic risk: up to 9 years of follow-up of 361,859 recipients. Pharmacoepidemiol Drug Saf 17: 27-36.
- Murtola TJ, Tammela TL, Määttänen L, Huhtala H, Platz EA, et al. (2010) Prostate cancer and PSA among statin users in the Finnish prostate cancer screening trial. Int J Cancer 127: 1650-1659.
- Sathyapalan T, Kilpatrick ES, Coady AM, Atkin SL (2009) The effect of atorvastatin in patients with polycystic ovary syndrome: a randomized doubleblind placebo-controlled study. J Clin Endocrinol Metab 94: 103-108.
- Banaszewska B, Pawelczyk L, Spaczynski RZ, Dziura J, Duleba AJ (2007) Effects of simvastatin and oral contraceptive agent on polycystic ovary syndrome: prospective, randomized, crossover trial. J Clin Endocrinol Metab 92: 456-461.
- Lemay A, Poulin Y (2002) Oral contraceptives as anti-androgenic treatment of acne. J Obstet Gynaecol Can 24: 559-567.
- Dahlgren E, Landin K, Krotkiewski M, Holm G, Janson PO (1998) Effects of two antiandrogen treatments on hirsutism and insulin sensitivity in women with polycystic ovary syndrome. Hum Reprod 13: 2706-2711
- 24. Pedersen C (2003) Lægemiddelkataloget. Dansk Lægemiddel Information A/S. Elanders Kungsbacka, Tryckeri, Sweden.
- 25. Harju M, Hamers T, Kamstra JH, Sonneveld E, Boon JP, et al. (2007) Quantitative structure-activity relationship modeling on in vitro endocrine effects and metabolic stability involving 26 selected brominated flame retardants. Environ Toxicol Chem 26: 816-826.

Page 10 of 10

- 26. Yang W, Mu Y, Giesy JP, Zhang A, Yu H (2009) Anti-androgen activity of polybrominated diphenyl ethers determined by comparative molecular similarity indices and molecular docking. Chemosphere 75: 1159-1164.
- Kovarich S, Papa E, Gramatica P (2011) QSAR classification models for the prediction of endocrine disrupting activity of brominated flame retardants. J Hazard Mater 190: 106-112.
- 28. Tamura H, Ishimoto Y, Fujikawa T, Aoyama H, Yoshikawa H, et al. (2006) Structural basis for androgen receptor agonists and antagonisms: Interaction of SPEED 98-listed chemicals and related compounds with the androgen receptor based on an in vitro receptor gene assay and 3D-QSAR. Bioorg Med Chem 14: 7160-7174.
- Hamers T, Kamstra JH, Sonneveld E, Murk AJ, Kester MH, et al. (2006) In vitro profiling of the endocrine-disrupting potency of brominated flame retardants. Toxicol Sci 92: 157-173.
- Xu LC, Liu L, Ren XM, Zhang MR, Cong N, et al. (2008) Evaluation of androgen receptor transcriptional activities of some pesticides in vitro. Toxicology 243: 59-65.
- Paris F, Balaguer P, Térouanne B, Servant N, Lacoste C, et al. (2002) Phenylphenols, biphenols, biphenol-A and 4-tert-octylphenol exhibit alpha and beta estrogen activities and antiandrogen activity in reporter cell lines. Mol Cell Endocrinol 193: 43-49.
- Tamura H, Maness SC, Reischmann K, Dorman DC, Gray LE, et al. (2001) Androgen receptor antagonism by the organophosphate insecticide fenitrothion. Toxicol Sci 60: 56-62.
- Sohoni P, Sumpter JP (1998) Several environmental oestrogens are also antiandrogens. J Endocrinol 158: 327-339.
- 34. Aït-Aïssa S, Laskowski S, Laville N, Porcher JM, Brion F (2010) Anti-androgenic activities of environmental pesticides in the MDA-kb2 reporter cell line. Toxicol In Vitro 24: 1979-1985.
- Shen O, Du G, Sun H, Wu W, Jiang Y, et al. (2009) Comparison of in vitro hormone activities of selected phthalates using reporter gene assays. Toxicol Lett 191: 9-14.
- 36. Katzenellenbogen JA, O'Malley BW, Katzenellenbogen BS (1996) Tripartite steroid hormone receptor pharmacology: interaction with multiple effector sites as a basis for the cell- and promoter-specific action of these hormones. Mol Endocrinol 10: 119-131.
- 37. Kojima H, lida M, Katsura E, Kanetoshi A, Hori Y, et al. (2003) Effects of a diphenyl ether-type herbicide, chlornitrofen, and its amino derivative on androgen and estrogen receptor activities. Environ Health Perspect 111: 497-502.
- 38. Multicase Inc (2011) Case Ultra User Guide 1.4.0.0.
- Satoh K, Nonaka R, Ohyama K, Nagai F, Ogata A, et al. (2008) Endocrine disruptive effects of chemicals eluted from nitrile-butadiene rubber gloves using reporter gene assay systems. Biol Pharm Bull 31: 375-379.
- 40. Takeuchi S, lida M, Kobayashi S, Jin K, Matsuda T, et al. (2005) Differential effects of phthalate esters on transcriptional activities via human estrogen receptors alpha and beta, and androgen receptor. Toxicology 210: 223-233.
- 41. Takeuchi S, Takahashi T, Sawada Y, lida M, Matsuda T, et al. (2009) Comparative study on the nuclear hormone receptor activity of various phytochemicals and their metabolites by reporter gene assays using Chinese hamster ovary cells. Biol Pharm Bull 32: 195-202.
- 42. Andersen HR, Vinggaard AM, Rasmussen TH, Gjermandsen IM, Bonefeld-Jørgensen EC (2002) Effects of currently used pesticides in assays for estrogenicity, androgenicity, and aromatase activity in vitro. Toxicol Appl Pharmacol 179: 1-12.
- 43. Kojima H, Takeuchi S, Uramaru N, Sugihara K, Yoshida T, et al. (2009) Nuclear hormone receptor activity of polybrominated diphenyl ethers and their hydroxylated and methoxylated metabolites in transactivation assays using Chinese hamster ovary cells. Environ Health Perspect 117: 1210-1218.

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- 44. Kawamura Y, Mutsuga M, Kato T, Iida M, Tanamoto K (2005) Estrogenic and Anti-Androgenic Activities of Benzophenones in Human Estroge and Androgen Receptor Mediated Mammalian Reporter Gene Assays. J Health Sci 51: 48-54.
- 45. Vinggaard AM, Joergensen EC, Larsen JC (1999) Rapid and sensitive reporter gene assays for detection of antiandrogenic and estrogenic effects of environmental chemicals. Toxicol Appl Pharmacol 155: 150-160.
- 46. Araki N, Ohno K, Takeyoshi M, Iida M (2005) Evaluation of a rapid in vitro androgen receptor transcriptional activation assay using AR-EcoScreen cells. Toxicol In Vitro 19: 335-352.
- 47. Satoh K, Ohyama K, Aoki N, Iida M, Nagai F (2004) Study on anti-androgenic effects of bisphenol a diglycidyl ether (BADGE), bisphenol F diglycidyl ether (BFDGE) and their derivatives using cells stably transfected with human androgen receptor, AR-EcoScreen. Food Chem Toxicol 42: 983-993.
- Vinggaard AM, Nellemann C, Dalgaard M, Jørgensen EB, Andersen HR (2002) Antiandrogenic effects in vitro and in vivo of the fungicide prochloraz. Toxicol Sci 69: 344-353.
- 49. Körner W, Vinggaard AM, Térouanne B, Ma R, Wieloch C, et al. (2004) Interlaboratory comparison of four in vitro assays for assessing androgenic and antiandrogenic activity of environmental chemicals. Environ Health Perspect 112: 695-702.
- Nikolov N, Grancharov V, Stoyanova G, Pavlov T, Mekenyan O (2006) Representation of chemical information in OASIS centralized 3D database for existing chemicals. J Chem Inf Model 46: 2537-2551.
- 51. Friedl H, Stampfer E (2006) Cross-validation. Encyclopedia of Environmetrics. Wiley Chichester, West Sussex, UK.
- Cooper JA 2nd, Saracci R, Cole P (1979) Describing the validity of carcinogen screening tests. Br J Cancer 39: 87-89.
- Ma R, Cotton B, Lichtensteiger W, Schlumpf M (2003) UV filters with antagonistic action at androgen receptors in the MDA-kb2 cell transcriptionalactivation assay. Toxicol Sci 74: 43-50.
- Tamura H, Yoshikawa H, Gaido KW, Ross SM, DeLisle RK, et al. (2003) Interaction of organophosphate pesticides and related compounds with the androgen receptor. Environ Health Perspect 111: 545-552.
- Orton F, Rosivatz E, Scholze M, Kortenkamp A (2011) Widely used pesticides with previously unknown endocrine activity revealed as in vitro antiandrogens. Environ Health Perspect 119: 794-800.
- Ibrahim NM, Marinovic AC, Price SR, Young LG, Fröhlich O (2000) Pitfall of an internal control plasmid: response of Renilla luciferase (pRL-TK) plasmid to dihydrotestosterone and dexamethasone. Biotechniques 29: 782-784.
- Auld DS, Southall NT, Jadhav A, Johnson RL, Diller DJ, et al. (2008) Characterization of chemical libraries for luciferase inhibitory activity. J Med Chem 51: 2372-2386.
- Auld DS, Thorne N, Nguyen DT, Inglese J (2008) A specific mechanism for nonspecific activation in reporter-gene assays. ACS Chem Biol 3: 463-470.
- 59. Daylight Theory Manual (2011) Daylight version 4.9. Daylight Chemical Information Systems, Inc. of Aliso Viejo, CA, USA.