

# Progesterone-induced Maturation and Down Regulation of Membrane Bound $\text{Na}^+$ , $\text{K}^+$ -ATPase

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## Abstract

Progesterone induces maturation by releasing oocyte from G2 of M1 cell cycle arrest. This process is rate limiting as it produces fertilizable eggs. Therefore, it draws a lot of attention.  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase is a membrane bound enzyme molecule that has known to have various functions. One of these functions is to act as receptor for Progesterone. In oocyte maturation to egg entirely depends on progesterone, a hormone that is known to reduce risk of cancer. Using EM Histo-cytochemical novel technique, we have shown that membrane bound  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase are gradually down-regulated following Progesterone-induced maturation. By Germinal Vesicle Break down,  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase is completely down-regulated from oolemma and only present in the narrow region of Germinal Vesicle Break Down. This is an important phenomena as this down-regulation coincides with cell entering M-phase. Here, I also briefly introduce you to a technique that only localizes phosphate cleaving membrane bound  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase.

**Keywords:**  $\text{Na}^+$ ;  $\text{K}^+$ -ATPase; Down regulation; Progesterone

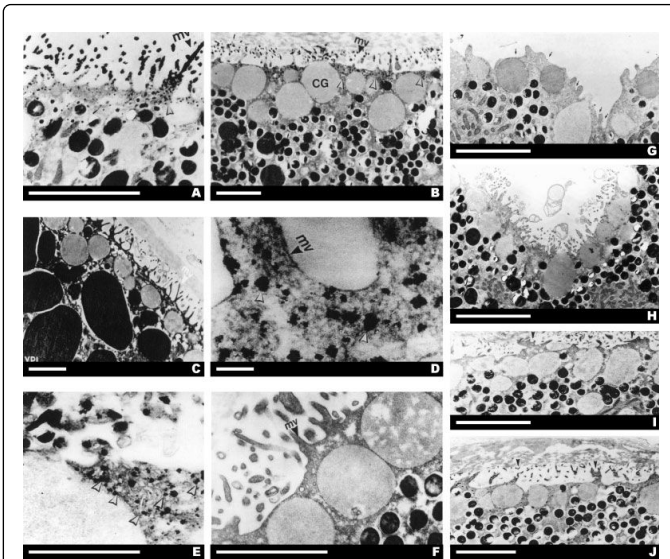
## Introduction

Progesterone induces fully grown Stage VI oocytes to mature that leads to Nuclear Envelope Break Down and the appearance of white spot in the animal hemisphere [1-3].  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase is a P-type ATPase that undergoes E1-E2 transition to translocate  $3\text{Na}^+$  outside cell against electrochemical gradient. In all living animal cells it regulates cell pH. It is also involved in firing electrical current in neuronal cells. It has specialized functions in muscles and cardiac cells. The molecular structure and genetic expression of this protein has been extensively studied [4,5]. It is a multigene protein. It is heterodimer of a catalytic  $\alpha$ -subunit of approximately 100-110 kD peptide and a  $\beta$ -subunit of 40 kD. Four isoforms of  $\alpha$ -subunit have been reported [4,6,7]. Three isoforms of  $\beta$ -subunit have also been identified [6]. The third subunit of  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase is  $\gamma$ -subunit of 7-10 kD [8,9]. Experimental evidences suggest that the  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity is modulated by protein kinase A and protein kinase C by phosphorylation [10,11].

$\text{Na}^+$ ,  $\text{K}^+$ -ATPase is a key instigator of cell polarity [12]. We have shown that  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity is asymmetrically localized over the animal hemisphere of fully differentiated Stage VI *Xenopus* oocytes [12]. The Vegetal pole that holds the vegetal cortices comprising VegT, TGF- $\beta$  and Cyclin-B lacks  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity completely. These Phosphate-cleaving membrane bound  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase sets up an internal gradient of  $\text{Na}^+$  that is critical in establishing polarity [13]. One important aspect of Progesterone-induced down-regulation of membrane bound  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase from animal hemisphere is down-regulation flattens the internal  $\text{Na}^+$  gradient. Therefore, it releases vegetal cortices from its position and moves upward following Progesterone-induced maturation [14] due to complete down-regulation of  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase from the animal hemisphere [12]. This down-regulation causes complete flattening of internal  $\text{Na}^+$  that is set due to asymmetric localization of  $\text{Na}^+$ ,  $\text{K}^+$ -

ATPase activity. This asymmetry sets up three internal  $\text{Na}^+$  gradients [12]. The vegetal hemisphere that lacks  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity has a sink for  $\text{Na}^+$ . The animal hemisphere that has  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity has medium level of  $\text{Na}^+$ . While the equatorial region has negligible to low  $\text{Na}^+$ . We believe, the acidic proteins such as Tgf- $\beta$  [15], Cyclin-B [16,17] and VegT [14] are localized in the vegetal cortices while the basic proteins such as AP-2 [18], Sox2 and Sox3 [19] are localized in the animal hemisphere and the neutral proteins such as Brachy [20] is localized in the equatorial region. Exceptions are there. As we know myoD is a basic protein and it is localized in the equatorial region possibly due to its binding partner. My estimation is  $\text{Na}^+$  is the key instigator of cell polarity. Therefore, this asymmetry is critical [12].

Following Progesterone-induced maturation,  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase is localized in the animal pole region. It appears that the Dorsal-ventral axis is already set up in the matured eggs with presence of ATPase activity towards the dorsal side (Figure 1) [12].  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase is key receptor molecule [1,2]. We believe, apart from Progesterone and Estradiol it also possibly act as receptor for sperm and we expect an up-regulation of ATPase activity at the sperm entry point. We think  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase may be the key receptor for hormones [1,2] and sperm. We are making greater stride to resolve the issue. As we know  $\text{Na}^+$  is the binding factor between sperm and egg therefore such asymmetric localization is critical. Surely this will be a turning point. Another important point about the down-regulation of  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase following Progesterone-induced maturation is arrest of cell cycle at M-phase. Does that mean  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity may have a role in metastasis? It would be interesting to observe  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity in uterine and cervical cancer by EM Histo-Cytochemical method, the technique that localizes phosphate cleaving  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase activity at the resolution of electron microscope. It may resolve what effect progesterone and estrogen have on uterine and cervical cancer (we are open to collaboration).



**Figure 1:** Localization of Na<sup>+</sup>, K<sup>+</sup>-ATPase activity following progesterone-induced maturation of stage-VI *Xenopus* oocytes. We see slight up-regulation (A) and then immediate complete down-regulation of Na<sup>+</sup>, K<sup>+</sup>-ATPase activity (B-H). A: An up-regulation of Na<sup>+</sup>, K<sup>+</sup>-ATPase activity within 1 hr after progesterone-induction. B: A substantial and near complete down-regulation of Na<sup>+</sup>, K<sup>+</sup>-ATPase activity following 3 hr of progesterone induction. C: Complete absence of Na<sup>+</sup>, K<sup>+</sup>-ATPase activity over the vegetal hemisphere. (D, E) Presence of vesicle associated reaction deposits in the sub cortical region of the oocyte following 3 hr of progesterone induction, suggesting endocytotic removal of the membrane Na<sup>+</sup>, K<sup>+</sup>-ATPase molecules from the plasma membrane. G: Presence of countable number of Na<sup>+</sup>, K<sup>+</sup>-ATPase molecules over the animal pole region of polar body extrusion following GVBD. Na<sup>+</sup>, K<sup>+</sup>-ATPase activity is completely down-regulated by GVBD, and vesicle-associated reaction deposits are absent from the cytoplasm of subcortical region. We believe the side that has maximum ATPase activity forms the dorsal side of the fertilized egg. This observation was only made possible by EM enzyme Histo-cytochemical technique: (I) Complete absence of reaction deposits, when K<sup>+</sup> is completely removed. (F, J) Complete absence and near complete absence of Na<sup>+</sup>, K<sup>+</sup>-ATPase activity over two different regions over the animal hemisphere in the presence of 10 μM Ouabain. (H) The presence of countable number of Ca<sup>2+</sup>, Mg<sup>2+</sup>-ATPase activity at the region of polar body extrusion over animal pole region following GVBD. Magnifications/bar sizes: (A) 15,000/1 mm; (B) 6,000/1 mm; (C) 5,000/1 mm; (D) 5,000/100 nm; (E) 30,000/1 mm; (F) 15,000/1 mm; (G-I) 10,000/1 mm; (J) 8,000/1 mm. (Adopted from Mohanty and Gupta, 2012).

Progesterone is a steroid hormone involved in female menstrual cycle, pregnancy (supports gestation) and embryogenesis of human and other species [20-22]. In clinical term, reduced progesterone adversely affects endometrium maturation and results in subfertility and pregnancy miscarriage [23,24]. On the contrary Estrogen exposure induces breast and endometrial cancer [25]. It also proliferated epithelial cells to develop into cancer [26]. Progesterone, on the otherhand, inhibits this estrogen-induced cell proliferation and stimulates epithelial differentiation [27]. Consequently, progesterone

is used therapeutically to inhibit the proliferation of estrogen-dependent endometrial cancers [28-30].

It is also important to know why a down-regulation is necessary before fertilization. The reason could be very rewarding. I believe the vegetal cortices released from vegetal hemispheres eggs following Progesterone-induced maturation has a significant role. Cyclin B moves upward and enters into the nucleus to complete division. Similarly, down-regulation makes eggs quiescent and prepares eggs to hostile environment and any damage to DNA. In *Xenopus*, eggs are laid in wet-land that remains hostile. We were the first to show a detailed analysis of Na<sup>+</sup>, K<sup>+</sup>-ATPase activity in any vertebrate oocytes and in maturing eggs [12] and the experiment using EM Histo-cytochemical technique was repeated 7 times with consistent results. In the process we realized Na<sup>+</sup>, K<sup>+</sup>-ATPase activity can be different if localized by EM Histo-cytochemical technique and immunocytochemical localization as later can localize epitope both from active and inactive Na<sup>+</sup>, K<sup>+</sup>-ATPase [31]. This protection allows organogenesis and patterning to proceed normally following fertilization to production of normal embryos.

EM Histo-cytochemical localization is the best technique to localize Na<sup>+</sup>, K<sup>+</sup>-ATPase activity. It gives most consistent results compared to immunocytochemical localization. The biochemical localization only shows presence of Na<sup>+</sup>, K<sup>+</sup>-ATPase activity but not their spatial distribution. For present study, we have used Mayhara et al. 1980 technique. It's a direct one-step method. In this technique we fixed the progesterone-induced and uninduced stage VI oocytes in 2% paraformaldehyde and 0.005% glutaraldehyde for 30 mins and then both groups of oocytes were processed as method completely described in Mohanty and Gupta [12]. The technique is so versatile that one can count individual phosphate cleaving ATPase molecules. We estimated up to 20 times higher ATPase activity over the animal hemisphere than the vegetal hemisphere.

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